Twenty-five Years Standardization and Quality Control of Nucleic Acid Amplification Technology for Detection of Blood Borne Viruses

Satellite Meeting before IPFA-PEI 25th Workshop on Surveillance and Screening of Blood Borne Pathogens, Tuesday, May 15th 2018, 15.00-18.00 hours at the Royal Olympic Hotel in Athens

14.30	**Welcome reception**	
	Presentation title	**Issues**
15.00	**Calibration of standards: foundation for understanding blood safety** Nico Lelie, Bio Quality Control, Heiloo, Netherlands	Calibration in IUs, copies, virions, minimum infectious dose, length of window periods, replacement of seroconversion panels by standard dilution panels. Alternatives to WHO standards for NAT validation
15.15	**External quality control ensuring sufficient analytical sensitivity of NAT assays** Nico Lelie, Bio Quality Control, Heiloo, Netherlands	Positioning of run controls in relation to NAT detection limits on native and inactivated standards
15.30	**Stability of native, lyophilized and inactivated standards** Harry van Drimmelen, Bio Quality Control, Heiloo, Netherlands	Long term stability of liquid frozen standards at -80˚C and -30 ˚C. Impact of lyophilization and inactivation on virus recovery and particle integrity. In use stability of QC samples at +4 ˚C and 21 ˚C
15.45	**The use of International Standards for quality control of NAT** Christoph Niederhauser, Berne, Blood Transfusion Center, Switzerland	Reduced NAT response on QC samples prepared from WHO replacement standards.
16.00	**Design and evaluation of multi-marker run control for cobas MPX assay** Marco Koppelman, Sanquin Blood Supply Foundation, Amsterdam, Netherlands	Analytical sensitivity of cobas MPX, Evaluation of two multi-marker run controls
16.15	**The use of ViraQ Check Controls for qualitative and quantitative NAT** Marion Vermeulen, South African National Blood Service, Johannesburg, RSA	Performance of run controls in Ultrio Plus/Elite and Abbott Real time assay. Quantification of NAT yield samples
16.30	**Monitoring analytical sensitivity of Procleix reagent batches with ViraQ Check and Trend Controls** Fiona Boland, Irish Blood Transfusion Center, Ireland	Is there variation in analytical sensitivity of TMA reagent batches or in HBV, HCV, HIV and HEV run control batches?
16.45	**Evaluation of external NAT controls from two manufacturers** Aneta Kopacz, Insitute of Hematology and Transfusion Medicine, Warsaw, Poland	Comparison of NAT response values on two run controls. Evaluation of quality aspects.
17.00	**Monitoring parvo B19V viral load in Procleix assays on Tigris and Panther platforms using ViraQ run control** Heli Tenkanen, Finish Red Cross Blood Service, Finland	A shift in quantitative values on B19V run control (10.000 IU/mL) after recalibration of TMA assay on WHO replacement standard
17.15	**Round Table Discussion** Chairman: Nico Lelie, Bio Quality Control, Netherlands	What are the essential elements for meaningful external quality control of NAT?
17.45	**Closing of Meeting**	

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