

# Calibration of standards: foundation for understanding blood safety



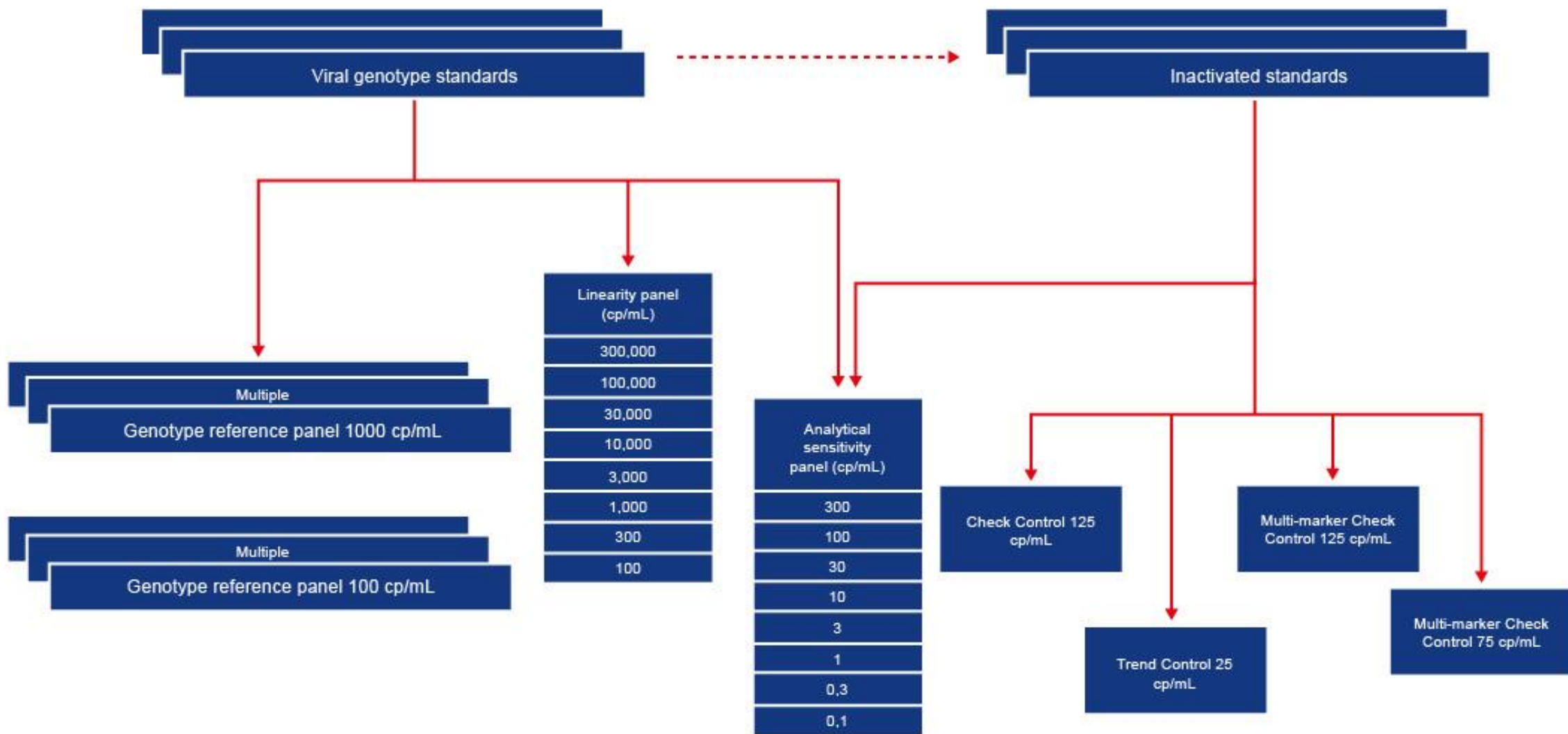
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Satellite Meeting before IPFA-PEI 25<sup>th</sup> Workshop Twenty-five Years Standardization and Quality Control of Nucleic Acid Amplification Technology for Detection of Blood Borne Viruses, May 15<sup>th</sup> 2018, Athens

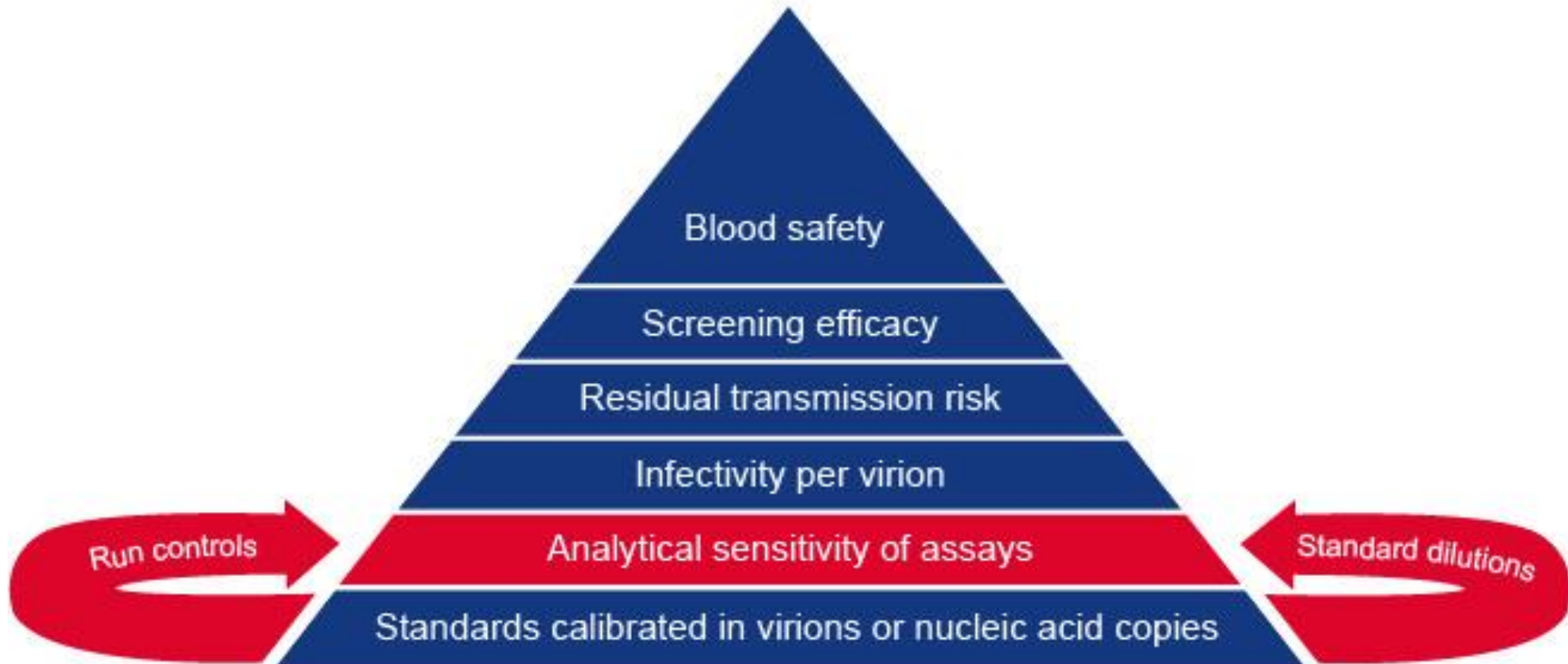


# Preparation of panels and controls for NAT since 1992<sup>#</sup>

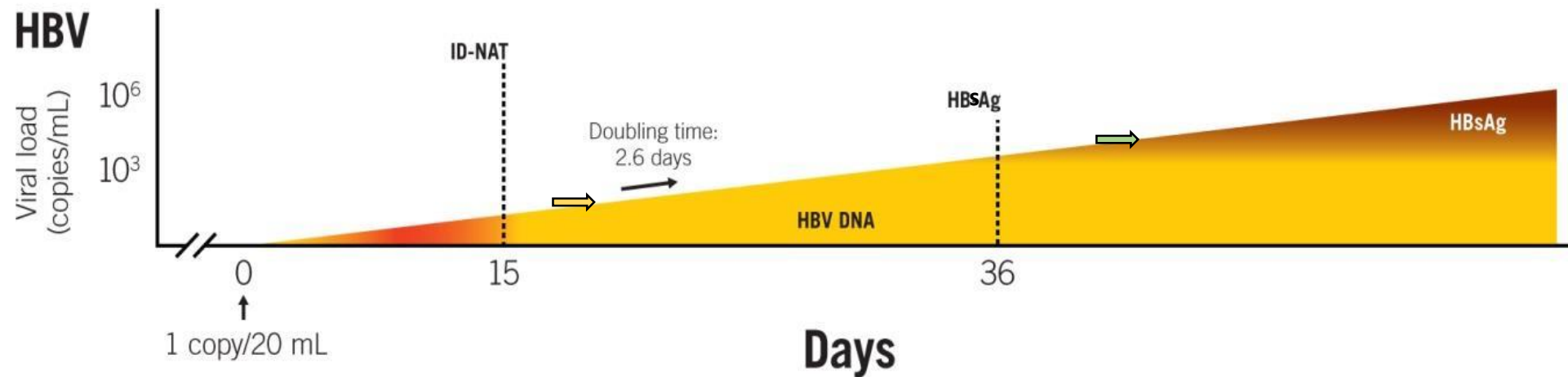
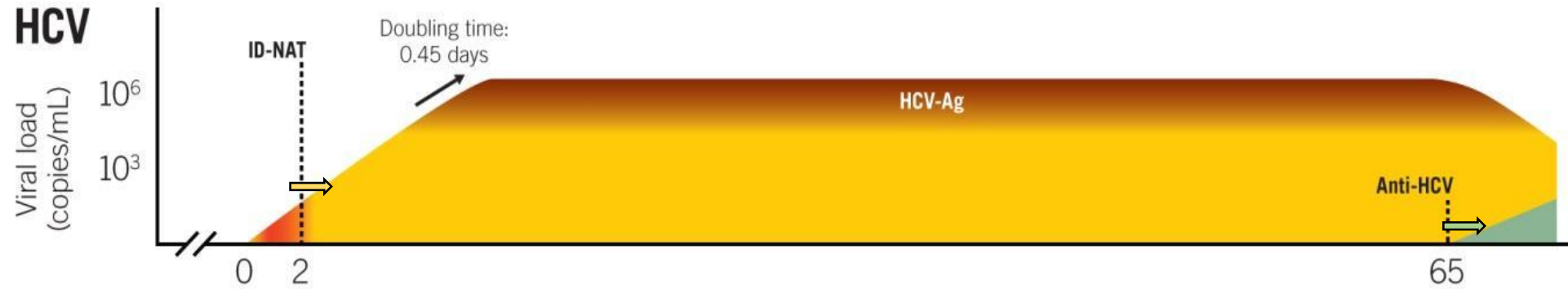
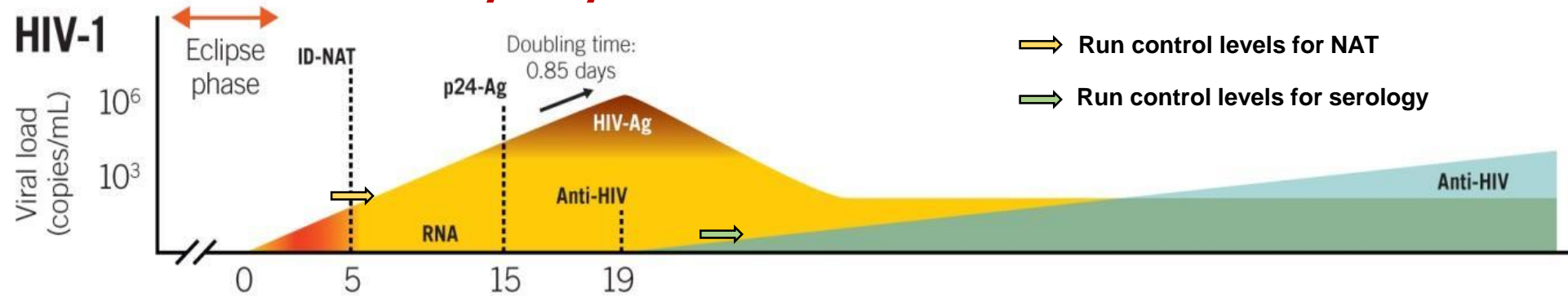


# First international proficiency study (Eurohep) published by Zaaijer et al (Lancet 1993;341:722-724)

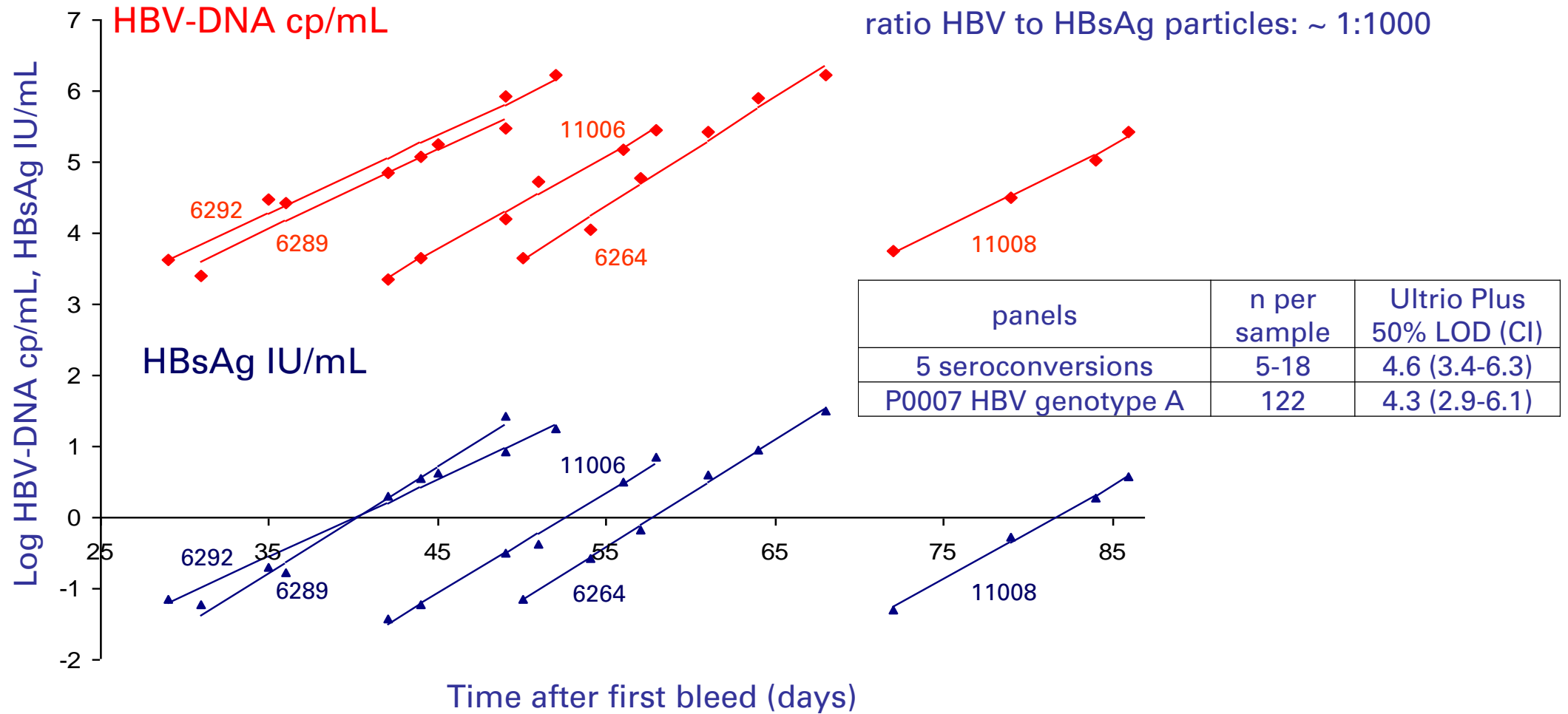
# Foundations for evaluating blood safety



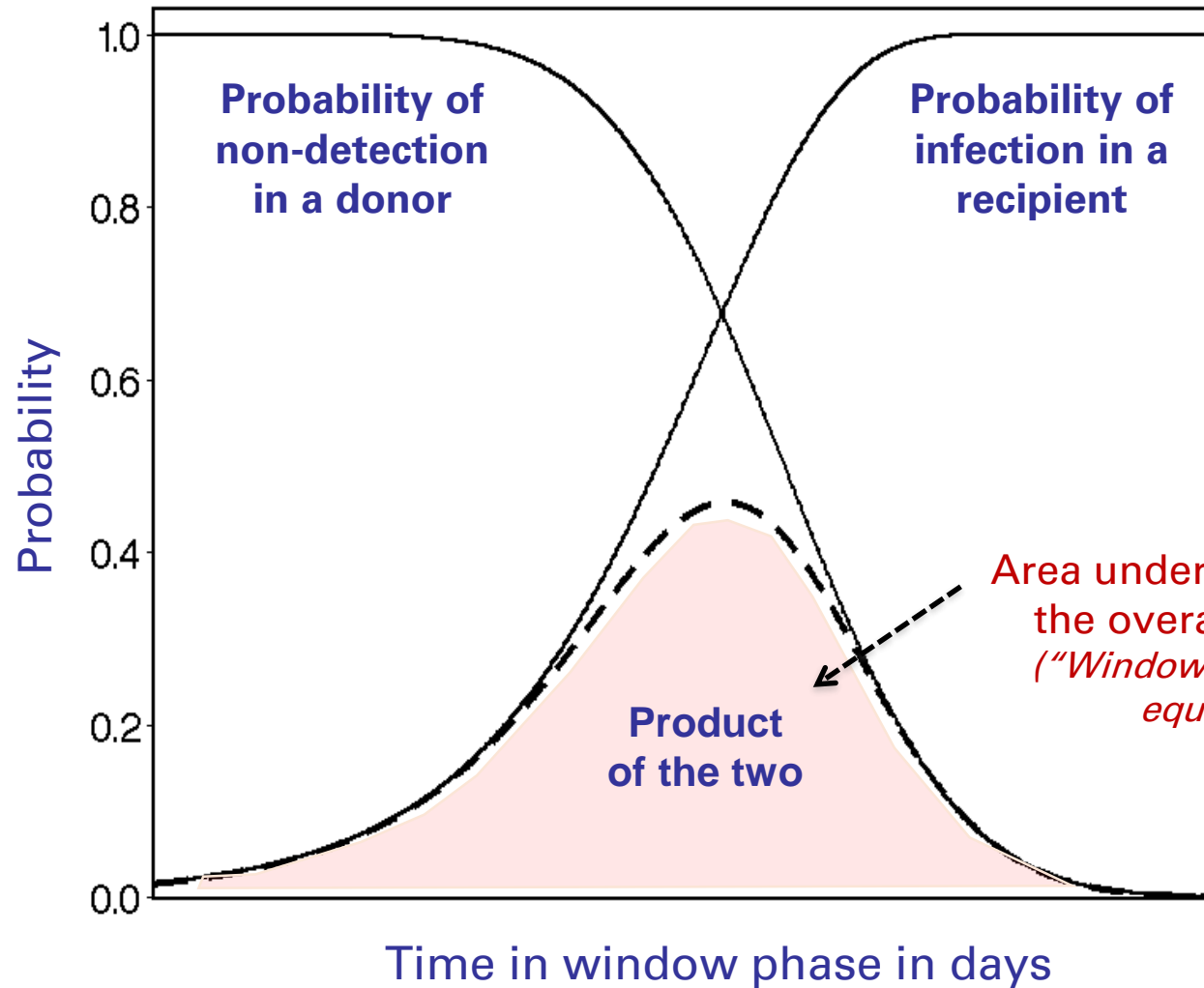
# Early dynamics of viral markers



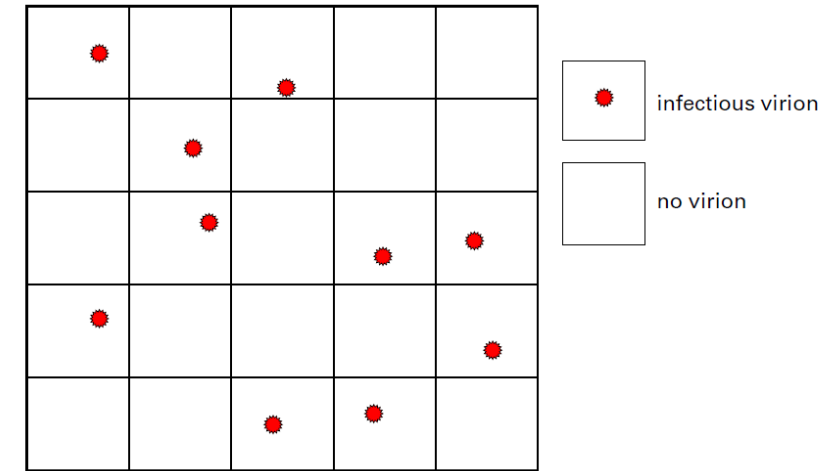
# Parallel kinetics of HBV-DNA and HBsAg in ramp up phase of 5 seroconversion panels



# Probability of infectivity during the window period



**Poisson distribution**  
*Probability of detection of virions in WP*



# Our view on metrological levels and traceability chain (ISO 17511:2003)^ for calibration of viral NAT standards in nucleic acid copies

Level	Traceable to SI unit	International reference measurement method	Calibrator material in measurement method	Reference standard
1	Yes	yes	NIST	NIST: P (Phosphate)
2	TBD	TBD: (Phosphate analysis, isotopic tracer, E260 and E280 extinction)#	P (Phosphate)	bDNA 3.0 assay calibrators (Purified in vitro RNA transcripts or DNA plasmids)#
3	No*	TBD: Multiple replicate bDNA 3.0 assays over time	bDNA assay calibrators	VQC-Sanquin standards
4	No	TBD: Separate calibration per NAT method in WHO collaborative study	VQC-Sanquin standard	WHO IS

^The IVD Directive refers to the ISO 17511:2003 standard for traceability of standards for IVDs and provides guidance on metrological levels

# Collins ML et al, Anal Biochem. 1995;226:120-9]

\* Extraction efficiency unknown



# Calibration of primary S0011 HBV genotype A standard in copies/mL by bDNA 3.0 assay as reference method<sup>^</sup>

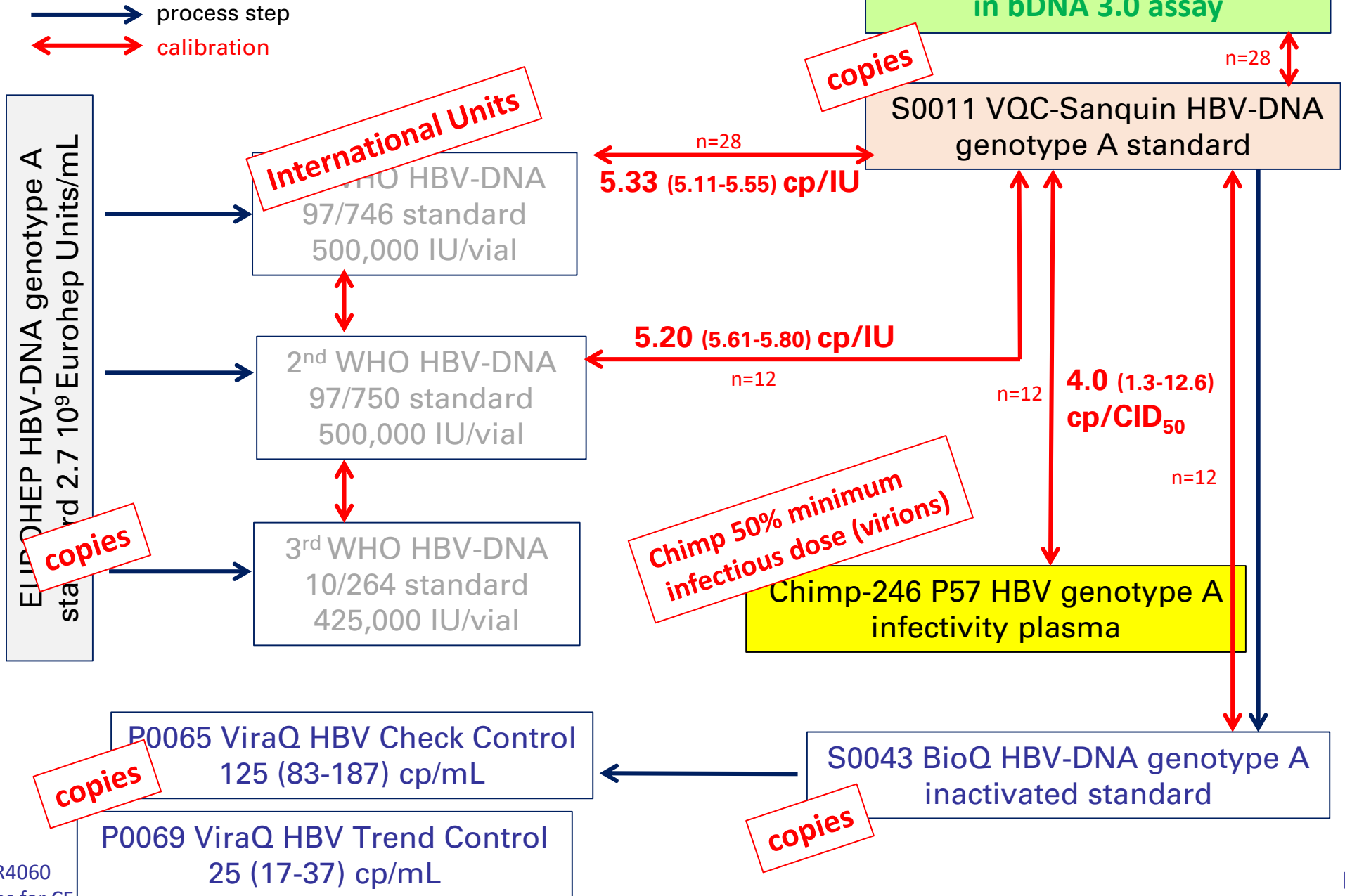
Assay	n S0011	copies/mL (95% CI) in VQC-Sanquin standard
Chiron bDNA 1.0	17	3.22 (3.13-3.32) x 10 <sup>9</sup>
<b>Siemens bDNA 3.0</b>	<b>28</b>	<b>2.15 (2.11-2.20) x 10<sup>9</sup></b>
Roche Amplicor Monitor	198	2.11 (2.05-2.17) x 10 <sup>9</sup>
Roche Taqman	8	2.38 (1.01-5.61) x 10 <sup>9</sup>
Digene HCS	42	1.63 (1.57-1.69) x 10 <sup>9</sup>

<sup>^</sup>Calibration of nucleid acid copies in bDNA assay is based on three physico-chemical techniques [Collins ML et al, Anal Biochem. 1995;226:120-9].

<sup>^</sup>Equivalent to copy numbers in Eurohep standard [Heermann KH et al, J Clin Microbiol. 1999;37:68-73 and Van Drimmelen et al, VR4060, BioQControl product files for CE marking]



# Calibration of HBV standards



# Calibration of S0011 HBV genotype A standard against 1<sup>st</sup> and 2<sup>nd</sup> WHO standard in bDNA 3.0 assay<sup>^</sup>

<b>International HBV Standard</b>	<b>VQC-Sanquin standard</b>	<b>n WHO</b>	<b>n S0011</b>	<b>copies/IU (95 %CI)</b>
WHO 97/746	S0011 HBV gt A	12	16	5.33 (5.11-5.55)
WHO 97/750	S0011 HBV gt A	6	6	5.20 (4.61-5.80)

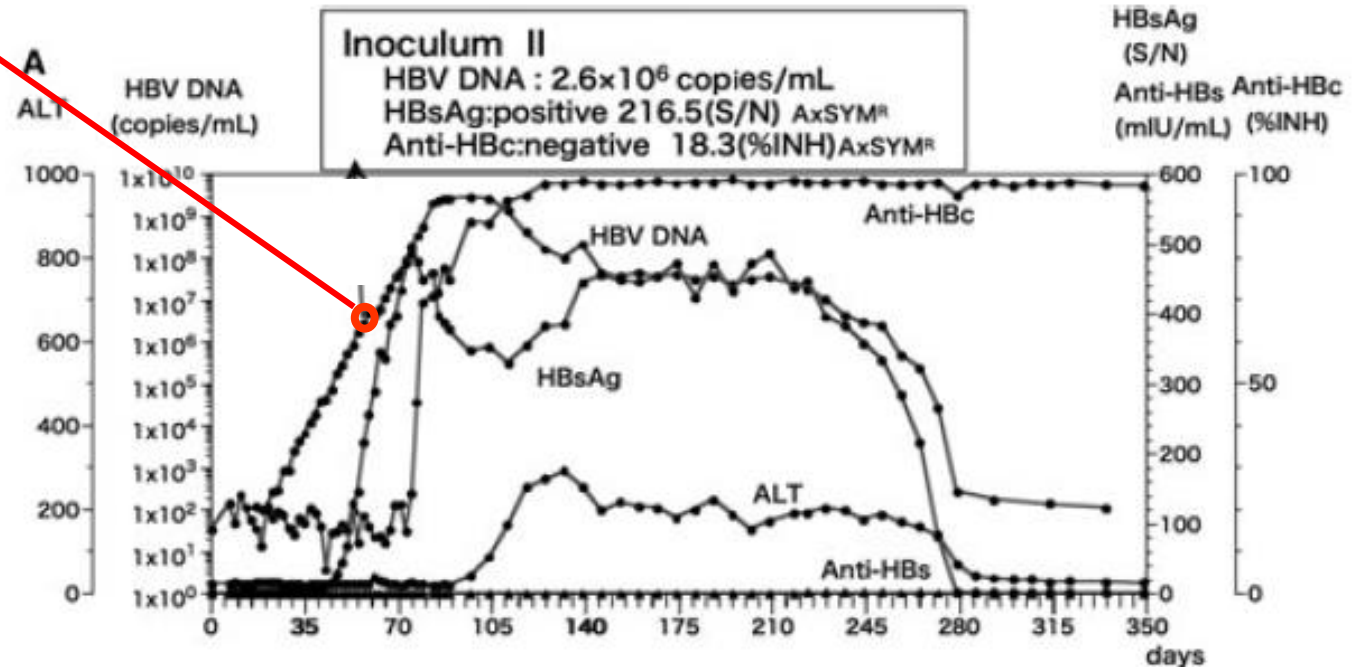
<sup>^</sup>Van Drimmelen et al, VR4060 BioQControl product files for CE registration

# Calibration of chimpanzee plasma of known infectivity against HBV genotype A standard in bDNA 3.0 assay

VQC-Sanquin standard	Chimp infectivity plasma <sup>^</sup>	n S0011	n Chimp	copies/CID <sub>50</sub> (range)
S0011 HBV gt A	C-246 (P-57) gt A	6	6	4.0 (1.3-12.6)

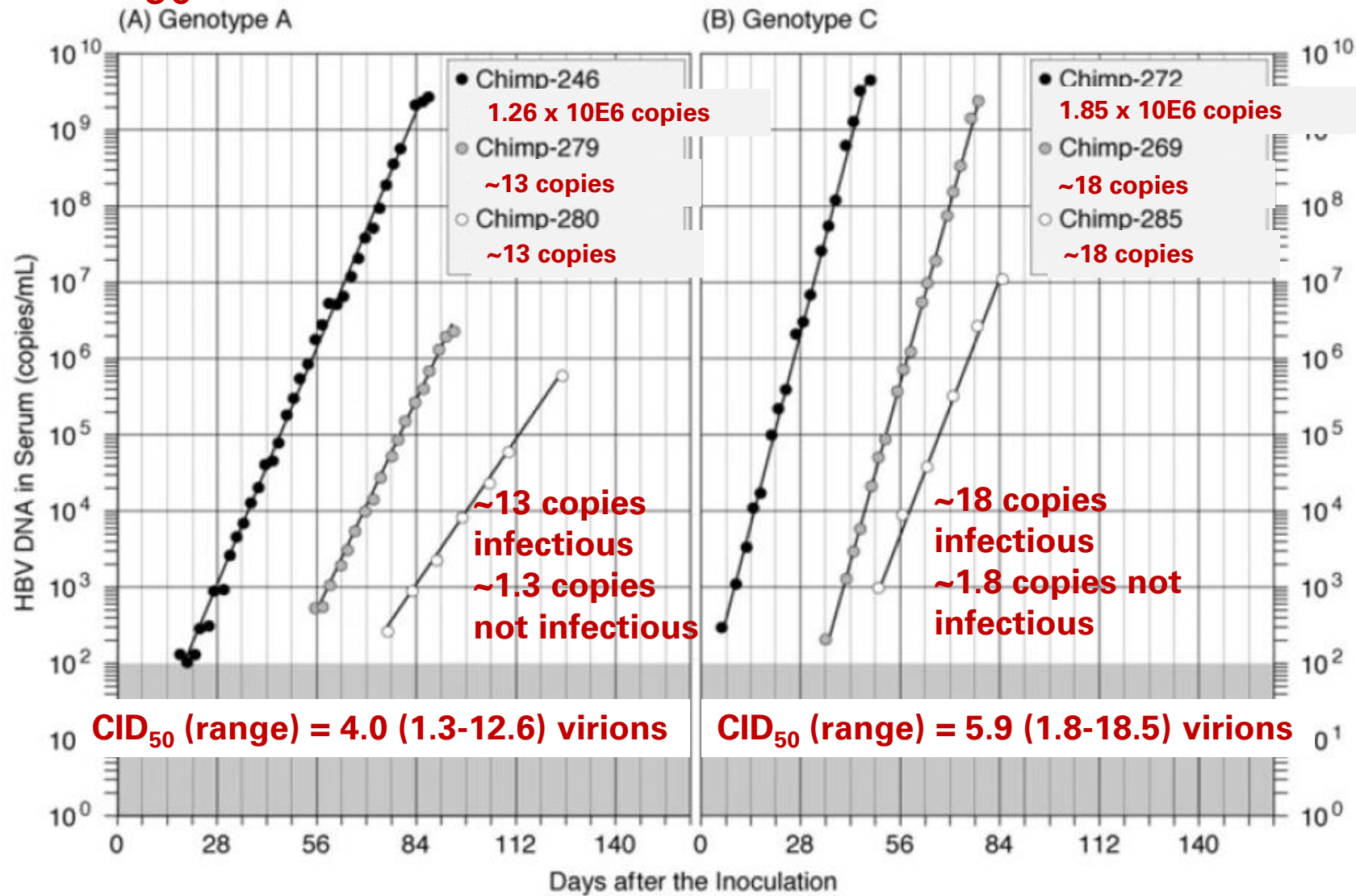
<sup>^</sup>Komiya K et al. Transfusion 2008;48:286-9

Van Drimmelen et al, VR4060  
(BioQControl CE files)



Sample C-246 (P57) was kindly provided by Prof Yoshizawa and Prof Tanaka, Hiroshima University, Japan

# Estimation of 50% chimpanzee minimum infectious dose (CID<sub>50</sub>) of HBV after recalibration of inocula



# 50% chimpanzee minimum infectious dose (CID<sub>50</sub>) of HCV after recalibration of inocula in bDNA 3.0 assay

<b>VQC-Sanquin standard</b>	<b>Chimp infectivity plasma<sup>^</sup></b>	<b>n C-210</b>	<b>n S0009</b>	<b>copies/ CID<sub>50</sub> (range)</b>
S0009 HCV gt 1	C-210 (wk-7) gt 1	6	6	8.1 (2.6-25.6)

<sup>^</sup>Katayama K et al. Intervirology 2004;47:57-64

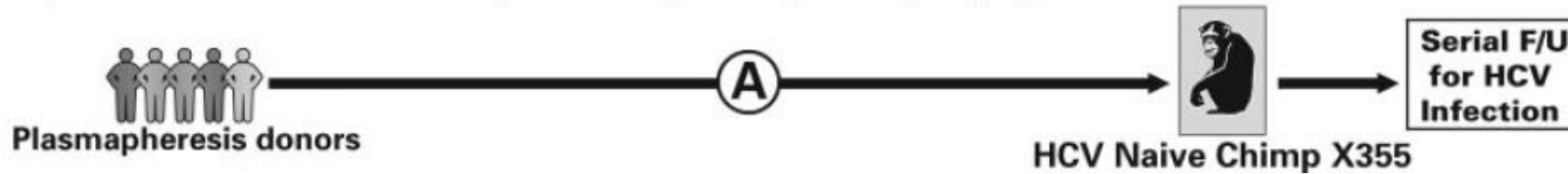
Van Drimmelen et al, VR4060 (BioQControl CE files)

Sample C-210 (wk-7) was kindly provided by Prof Yoshizawa and Prof Tanaka, Hiroshima University, Japan

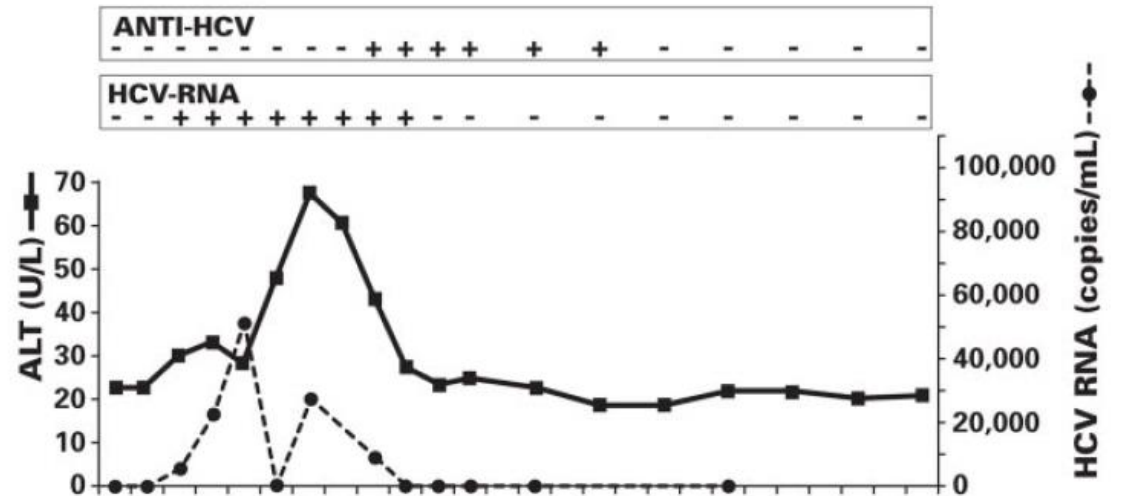
# Re-estimation of infectious dose of HCV in very early ramp-up plasma

Busch et al. Blood 2012;119(26):6326-6334

## Experiment I (Infectivity of very early ramp-up plasma)



HCV-RNA estimation in 50 mL inoculum of infectious donor by probit analysis (2/27 TMA reactive)	
according to Busch et al [Blood 2012:119:6326]	on S0009 VQC-Sanquin standard dilutions
1.2 (0.5-1.9) cp/mL 60 (25-95) copies	0.3 (0.1-0.6) cp/mL 14 (5-30) copies



**CID<sub>50</sub> (range) could be as low as 4.4 (1.4-14) virions**

For worst case risk modelling according to Weusten et al [Transfusion 2011;51:203-15] a MID<sub>50</sub> (range) of 3.2 (1-10) copies or virions was proposed by Kleinman et al [Transfusion 2009;49:2454-89]

## Current NAT efficiencies<sup>^</sup> estimated on primary Eurohep and VQC-Sanquin standards calibrated in copies/mL

Standard	Assay	n	% efficiency (CI)
S0011 VQC-Sanquin HBV-DNA genotype A2	Ultrio Plus	48	27 (20-36)%
	Ultrio Elite	74	35 (23-53)%
S0010 Eurohep HBV-DNA genotype A2	Ultrio Plus	96	34 (27-43)%
	cobas MPX	48	42 (25-68)%
S0009 VQC-Sanquin HCV-RNA genotype 1	Ultrio Plus	48	74 (53-99)%
	Ultrio Elite	112	81 (67-97)%
	cobas MPX	60	24 (18-30)%
S0012 VQC-Sanquin HIV-1 RNA subtype B	Ultrio Plus	48	75 (54-101)%
	Ultrio Elite	24	71 (47-100)%
	cobas MPX	48	55 (42-73)%

<sup>^</sup>calculated from 63% LOD (CI) in cp/mL on standard dilution panels  
 0.5 mL plasma input in Ultrio versions  
 1.0 mL plasma input in cobas MPX.



# Calibration of VQC-Sanquin HIV-RNA subtype B standard on 1<sup>st</sup> and 2<sup>nd</sup> IS in WHO collaborative study

Assay	N assays			VQC cp/IU on 1 <sup>st</sup> WHO (97/656) standard		VQC cp/IU on 2 <sup>nd</sup> WHO (97/650) standard	
	1 <sup>st</sup> WHO	2 <sup>nd</sup> WHO	VQC	mean	(95%CI)	mean	(95%CI)
Abbott LCx	14	15	14	0.76	(0.60-0.96)	0.69	(0.56-0.86)
Roche Amplicor Monitor	125	134	112	0.70	(0.60-0.81)	0.93	(0.80-1.08)
<b>Siemens bDNA 3.0</b>	<b>64</b>	<b>69</b>	<b>48</b>	<b>0.39</b>	<b>(0.34-0.44)</b>	<b>0.58</b>	<b>(0.51-0.66)</b>
Organon Teknika NucliSens <sup>^</sup>	46	51	36	0.80	(0.69-0.92)	0.43	(0.36-0.50)
Roche Amplicor Mon UltraSens	16	15	11	0.51	(0.27-0.95)	0.86	(0.49-1.51)

calculated from raw data reported by the laboratories participating in the first WHO collaborative study (Holmes H et al, J. Virological Methods 2001, 92; 141-150)

<sup>^</sup> primer mismatch

# Comparable NAT detection limits on primary and secondary HBV standards

HBV genotype A standard	NAT method	n	50% LOD (CI) cp/mL	95% LOD (CI) cp/mL
WHO 97/750	Ultrio Plus	303	4.4 (3.3-5.9)	28.4 (18.0-57.7)
S0011 VQC-Sanquin	Ultrio Plus	48	4.8 (3.7-6.2)	38.8 (25.6-68.5)
S0010 Eurohep	Ultrio Plus	96	3.6 (2.9-4.4)	40.4 (29.2-60.2)
WHO 97/750	Ultrio Elite	252	4.4 (3.6-5.4)	30.9 (22.4-47.4)
S0011 VQC-Sanquin	Ultrio Elite	74	3.4 (2.3-4.8)	43.2 (24.8-98.0)
S0010 Eurohep	Ultrio Elite	24	7.9 (5.5-11.2)	49.1 (29.4-116)
WHO 97/750	cobas MPX	12	1.8 (0.93-2.8)	8.0 (4.4-37.4)
S0011 VQC-Sanquin	cobas MPX	24	1.9 (1.3-2.7)	13.0 (7.7-29.6)
S0010 Eurohep	cobas MPX	48	1.7 (1.0-2.4)	10.3 (6.2-28.8)

1 IU = 5.33 (5.11-5.55) copies

# Comparable NAT detection limits on primary and secondary HIV-1 standards

HIV-1 subtype B standard	NAT method	n	50% LOD (CI) cp/mL	95% LOD (CI) cp/mL
WHO 97/650	Ultrio Plus	288	2.4 (2.2-2.6)	13.4 (11.4-16.3)
S0012 VQC-Sanquin	Ultrio Plus	48	1.7 (1.3-2.2)	15.1 (9.9-26.9)
WHO 97/650	Ultrio Elite	229	2.2 (1.4-3.2)	17.2 (10.3-40.1)
S0012 VQC-Sanquin	Ultrio Elite	24	2.1 (1.5-2.9)	9.0 (5.8-19.5)
WHO 97/650	cobas MPX	12	2.7 (1.7-3.9)	5.8 (3.9-24.9)
S0012 VQC-Sanquin	cobas MPX	48	1.3 (1.0-1.6)	7.3 (5.3-11.8)

1 IU = 0.58 (0.51-0.66) copies

# Comparable NAT detection limits on primary and secondary HCV standards

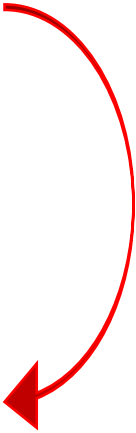
HCV genotype 1 standard	NAT method	n	50% LOD (CI) cp/mL	95% LOD (CI) cp/mL
WHO 96/798	Ultrio	24	2.6 (1.9-3.4)	20.5 (13.9-33.6)
WHO 06/100	Ultrio	32	2.5 (1.8-3.4)	18.9 (11.7-39.0)
S0009 VQC-Sanquin	Ultrio	36	2.9 (2.1-3.9)	23.9 (15.0-46.7)
WHO 06/100	Ultrio Plus	288	2.9 (2.0-4.2)	20.7 (12.2-50.3)
S0009 VQC-Sanquin	Ultrio Plus	48	1.8 (1.3-2.3)	15.1 (9.9-26.6)
WHO 06/100	Ultrio Elite	244	3.4 (2.0-5.4)§	26.8 (14.2-89.4)§
S0009 VQC-Sanquin	Ultrio Elite	112	1.7 (1.5-2.0)§	10.0 (7.7-13.8)§
S0009 VQC-Sanquin	cobas MPX	60	2.9 (2.3-3.6)	20.4 (14.3-33.3)

1 IU = 2.73 (1.4-4.8) copies

§ p<0.05

S0009 VQC-Sanquin HCV standard is currently used as alternative to WHO 06/100 and 06/102 standards in analytical sensitivity studies

50% LOD (CI) IU/mL	95% LOD (CI) IU/mL
0.27 (0.73 -1.98)§	9.82 (5.20-32.7)§
0.37 (0.55-0.73)§	3.66 (2.82-5.05)§



# Conclusions

- Calibration of VQC-Sanquin HBV, HCV and HIV-1 standards in nucleic acid copy (or virion) numbers enabled estimating lengths of diagnostic window periods and residual transfusion transmission risk.
- The likelihood of nearly accurate calibration of primary VQC-Sanquin standards in copies/mL with bDNA 3.0 reference assay has been supported by our studies with chimpanzee plasmas of known infectivity and by limiting dilution NAT efficiency studies.
- Viral standard dilution panels calibrated in copies/mL can replace seroconversion panels for evaluating sensitivity of NAT (and antigen) assays and estimating diagnostic window periods.
- VQC-Sanquin HBV, HCV, HIV-1, HAV, Parvo B19V standards are directly traceable to 1<sup>st</sup> and 2<sup>nd</sup> WHO standards and can be used as an alternative to WHO standards in analytical sensitivity studies of NAT methods.
- The amount of HIV-1 copies per IU increased approximately 1.5 fold by replacing the 1<sup>st</sup> by the 2<sup>nd</sup> WHO IS (because of variable detection efficiency of NAT methods for the two International Standards in the WHO collaborative study).