



# **P0064**

## **ViraQ HIV-1 Check 125**



The kit insert contains a detailed protocol and should be read carefully before testing the run control to ensure optimal performance



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## Intended Use

P0064 ViraQ HIV-1 Check 125 is intended to be used as external run control for human immunodeficiency virus type 1 (HIV-1) RNA amplification tests in combination with the assays on the plat-forms defined in Table 1. The run control helps laboratories to ensure sufficient analytical sensitivity and consistent performance of:















- qualitative multiplex nucleic acid amplification tests (NAT) for blood screening
- quantitative NAT methods with a lower limit of quantification (LOQ) sufficiently below the run control concentration of 125 copies/mL (~ 215 International Units (IU)/mL)

**Table 1.** Assays and platforms covered by P0064 ViraQ HIV-1 Check 125 run control

Assays (manufacturer)	Platform	Test environment
Procleix Ultrio® (Grifols)	Procleix Tigris®	Blood screening
Procleix Ultrio Plus® (Grifols)		
Procleix Ultrio Elite® (Grifols)	Procleix Panther®	Viral load monitoring
Aptima® HIV-1 Quant Dx (Hologic)	Panther®	

P0064 ViraQ HIV-1 Check 125 should not be used to replace the internal controls or calibrators in the test kits. The test result on the run control should not be used to reject the run or delay the release of test results on donor or patient samples.

## Key to Symbols Used

						
Manufacturer	Lot number	Catalogue number	In vitro diagnostic medical device	Country of origin	Contains human blood derivatives	Store below -30°C
						
Expiry date	Number of controls	Caution	CE mark with Notified Body number	Unique device identification	Read instructions for use	Biological substance Category B

## Principle of method

P0064 ViraQ HIV-1 Check 125 control has been formulated to mimic natural plasma specimens with a low HIV-1-RNA concentration. After thawing the run control tubes are ready for use and can be placed at random positions in sample racks on the NAT platforms. The run control contains 125 copies/mL of HIV-1 RNA (equivalent to 215 IU/mL) and has been designed to ensure sufficient analytical sensitivity of transcription mediated amplification (TMA) tests in blood screening laboratories. The run control is also suitable for monitoring performance of quantitative HIV-1 RNA assays in diagnostic laboratories using real time TMA or polymerase chain reaction (PCR) methods. The HIV-1-RNA concentration in the run control has been set at ~6 times the 95% lower limit of detection (LOD) of the Ultrio (Plus and Elite) assays (table 2)<sup>1-5</sup> and at 3-5 times the LOQ of the above mentioned quantitative NAT assays<sup>5</sup>. The positioning of P0064 ViraQ HIV-1 Check 125 control ensures reactivity rates above 99.5% in the NAT systems listed in table 1. The run control enables laboratories to be alerted in case of a significant reduction of analytical sensitivity of NAT test systems and to identify changes in the (precision of) viral load tests

over time. The run control is a dilution of the S0041 inactivated HIV-1-RNA subtype B standard, prepared by heat-inactivation of tissue culture derived virus spiked in plasma<sup>6-8</sup>. The plasma matrix in which the run control is diluted is manufactured from plasma units that tested negative for all relevant markers of blood borne viruses. The S0041 HIV-1 standard has been calibrated in copies/mL and IU/mL against the Viral Quality Control (VQC)-Sanquin and World Health Organization (WHO) International Standards (figure 1). The low concentration of HIV-1 subtype B in the run control is meant to be representative for HIV-1 subtypes A-K and circulating recombinant forms that are prevalent in different geographical regions of the world (although not all genetic variants are detected with comparable analytical sensitivity by the commercial NAT assays)<sup>4,9-11</sup>. A positive (and quantifiable) result on the run control indicates that the NAT method has been performed with sufficient analytical sensitivity. A non-reactive result or a reactive result below the LOQ is indicative of reduced analytical sensitivity of the NAT system and should trigger investigation of the technical performance of the assay. The run control generates sample to cut-off (S/CO) ratios in the Procleix Ultrio assay versions and Ct values or viral loads (expressed in copies/mL) in quantitative TMA and real time PCR assays. Statistical analysis of these assay response values generated over a certain period of time allows for comparison of analytical performance of NAT reagent batches and laboratory instruments.

**Table 2.** Detection limits on native and inactivated HIV-1 standard dilution panels in Procleix Ultrio assay versions

standard	panel	NAT method	n	50% LOD (CI) cp/mL	95% LOD (CI) cp/mL
S0041 BioQ HIV-1 RNA subype B inact.	P0026	Ultrio	52	3.1 (2.4-3.9)	20.2 (13.9-33.3)
S0012 VQC-Sanquin HIV-1 RNA subtype B	P0025	Ultrio	60	1.5 (1.0-2.2)	11.2 (6.3-29.8)
	P0025	Ultrio Plus	48	1.7 (1.3-2.2)	15.1 (9.9-26.9)
	P0025	Ultrio Elite	24	2.1 (1.5-2.9)	9.0 (5.8-19.5)
WHO HIV-1 RNA 97/650 <sup>#</sup>	P0022	Ultrio	40	2.6 (2.1-3.3)	11.8 (8.2-20.7)
	P0022	Ultrio Plus	288	2.4 (2.2-2.6)	13.4 (11.4-16.3)
	P0022	Ultrio Elite	229	2.2 (1.4-3.2)	17.2 (10.3-40.1)

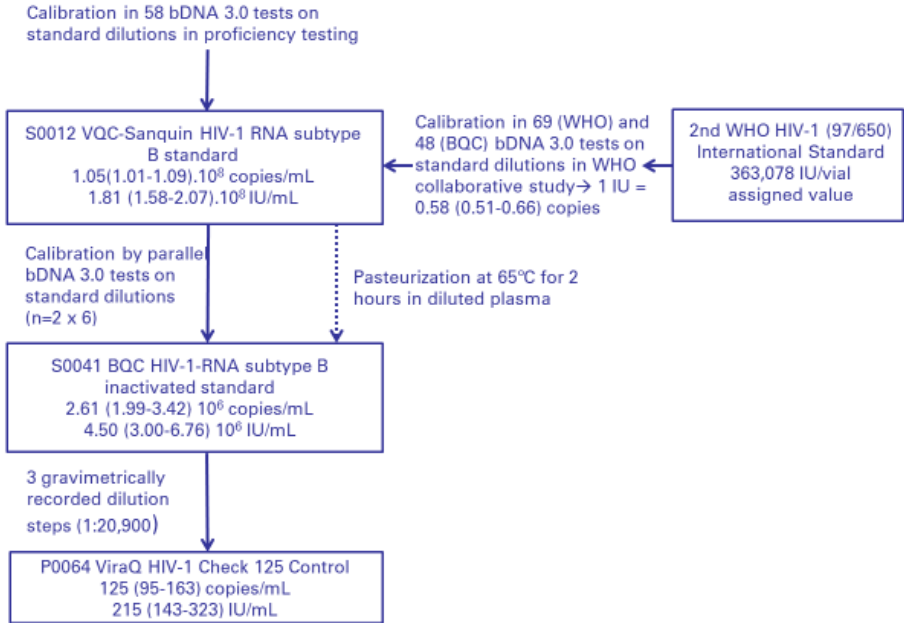
# 1 IU = 0.58 copy

### Traceability to HIV-1 RNA copies and International Units

Figure 1 shows the traceability chain between the ViraQ run control, the Bio Quality Control (BQC) standard, VQC-Sanquin standard and the 2<sup>nd</sup> WHO 97/650 International Standard for HIV-1 RNA. The inactivated S0041 HIV-1 RNA standard (used for preparation of the P0064 ViraQ run control) has been calibrated in copies/mL by replicate testing in the Siemens Versant bDNA 3.0 assay<sup>12</sup> against the historically established S0012 VQC-Sanquin HIV-1 RNA subtype B standard<sup>13</sup>. The VQC-Sanquin HIV-1 RNA subtype B standard has been calibrated at 0.58 (0.51-0.66) copies per IU against the second WHO HIV-1 RNA 97/650 standard in multiple replicate bDNA 3.0 assays<sup>14-16</sup>. It must be emphasized that this conversion factor from copies to IU values was 0.39 (0.34-0.44) for the 1<sup>st</sup> WHO HIV-1 97/656 standard and has not been determined for the 3<sup>rd</sup> WHO 10/152 replacement standard<sup>14</sup>. The accurate calibration of the native VQC-Sanquin and inactivated BQC HIV-1 subtype B standards in copies/mL and IU/mL has been confirmed in analytical sensitivity studies of the Grifols Procleix TMA and Roche cobas MPX assays<sup>4,14</sup>. The BQC manufacturing and quality control procedures guarantee consistent virus concentrations

in consecutive ViraQ HIV-1 Check 125 batches<sup>17</sup>. The inactivated BQC HIV-1 subtype B standard is available in sufficient supply to ensure batch consistency of ViraQ run controls for a prolonged period of time.

**Figure 1.** Traceability chain between run control, BQC and VQC-Sanquin standards and WHO International Standard



**Stability of HIV-1 standards and run control**

The long term stability of the liquid frozen S0041 HIV-1 standard stored below -65°C has been firmly established<sup>18</sup>; hence the stock solution from which the run control is prepared has shown to be stable in the BQC storage facilities. Real time stability experiments using quantitative NAT assays showed less than 10% degradation of HIV-1 RNA per year in P0064 ViraQ HIV-1 Check 125 control (and in standard dilutions of higher concentration) when stored at -30°C<sup>18</sup>. Hence, it can be guaranteed that the run control is still functional and should generate a reactivity rate greater than 99.5% when stored at -30°C and used before the expiration date (two years after preparation of the run control batch)<sup>17,18</sup>.

**Kit Contents (materials provided)**

The run control contains human plasma without preservatives and is provided in three formats as detailed in Table 3.

**Table 3. Description of kit formats and contents**

Cat. Code	UDI code	Quantity run control	Size vials	packing
P0064/01	8718719830640	60 x 1.5 mL	10 mL	60 vials in rack/box
P0064/02	8718719830283	10 x 1.5 mL	10 mL	Plastic zip bag
P0064/03	8718719830284	10 x 1.5 mL	2 mL	Plastic zip bag

P0064/01 and P0064/02 are intended to accommodate both blood screening and diagnostic laboratories. To facilitate automation the run control is presented in a polypropylene tube with screw cap comparable in size to vacutainer tubes used for donor sample collection. The tube label has a barcode identifying the product, sequential batch number and marker HIV-1. The barcode can be read by the automated NAT systems.

P0064/03 is intended to accommodate molecular diagnostic laboratories using smaller vials in routine procedures. The vial label does *not* have a barcode; the control should be identified on the work list.

### **Materials required but not supplied**

The test kits and liquid handling devices provided by the NAT manufacturer as specified in Table 1.

### **Storage instructions**

The run controls should be stored at or below -30°C for a maximum of two years<sup>18</sup>. Once thawed the run control samples should be used within 8 hours. During this period, when not in use, store sample at 2-8°C<sup>18</sup>. Do not refreeze the controls after thawing to prevent formation of cryoprecipitates. Any control sample that appears cloudy or contains precipitates after thawing and mixing should be discarded.

### **Warning and precautions**

Although P0064 ViraQ HIV-1 Check 125 contains inactivated HIV-1 particles<sup>6-8</sup> the plasma may still be potentially bio-hazardous. The matrix is prepared from human blood plasma that tested negative for blood borne viruses (HBV-DNA, HCV-RNA, HIV-RNA, HBsAg, anti-HBc, anti-HIV, anti-HCV and anti-Treponema *pallidum*). No test method can offer complete assurance that products derived from human blood cannot transmit (unknown) infectious agents. The run control should only be used by trained laboratory workers who are aware of the potential risk of infectious agents in human plasma samples and take the necessary precautions. Observe the universal precautions for prevention of transmission of infectious agents when handling these materials<sup>19,20</sup>.

- Do not pipette by mouth.
- Use personal protective equipment, including lab coats, gloves and safety glasses.
- Do not eat, drink or smoke in areas where the run control is handled.
- Disinfect spills using a 0.5% hypochlorite solution (1:10 v/v household bleach) or equivalent disinfectant.
- Dispose unused or spilled materials according to the normal practices for biological waste disposal in your institution.
- If precipitates are visible, mix the run controls for 2 minutes thoroughly.
- Once thawed, do not re-freeze and thaw the run control samples to avoid formation of cryoprecipitates that could alter reactivity or cause pipetting errors in the automated sampling systems.
- Store run controls in an upright position.

## Reagent preparation

- Thaw the run control quickly in a water bath at 37°C.
- Mix gently during thawing until contents are just thawed.
- Immediately after thawing remove the run control tube from the water bath.
- Vortex the run control.
- Give a short spin in a centrifuge to remove liquid before releasing screw cap from vial.
- Minimise the time period from thawing until usage of the control samples.
- Use within 8 hours after thawing
- After thawing when not in use: store at 2-8°C

## Test procedure and calculations

The run control should be tested in a manner identical to that of clinical specimens and the result be calculated according to the instructions for use of the NAT procedure.

The following sections in this package insert provide guidance on interpretation and analysis of test results on P0064 ViraQ HIV-1 Check 125. The statistical evaluation methods were developed by BioQControl and were not reviewed nor approved by the manufacturer of the Ultrio assay versions.

### *Qualitative detection of HIV-1 RNA in Procleix Ultrio versions*

The results of the Procleix Ultrio, Ultrio Plus and Ultrio Elite assays are expressed as a sample to cut-off ratio (S/CO). P0064 ViraQ HIV-1 Check 125 Control should react positive in more than 99.5% of TMA test runs.

More than 99% of test results on the run control are expected in the (near) saturated range of the TMA assay with S/CO values equal to or above 8.0 (range 8.0-15.0). Less than 1% of results are expected in the lower dynamic range of the TMA assay with S/CO ratios below 8.0 (see interpretation of test results below)<sup>17</sup>. A Levey-Jennings QC chart can be used to monitor the performance of the Ultrio assay versions on the run control.

### Levey-Jennings QC chart.

The S/CO responses on ViraQ HIV-1 Check 125 in the Ultrio Plus and Elite assay versions are not normally distributed, also not after transformation of the S/CO ratios. For developing a Levey-Jennings QC chart a distribution-free approach can be taken whereby the ranges containing 95% and 99% of the data are calculated. For this purpose the 0.5% and 99.5% percentiles are calculated for the 99% interval, and the 2.5% and 97.5% percentiles for the 95% interval. In case the total dataset does not contain sufficient data, values just outside the observed range can be presented in order to allow for a graphical presentation. For this non-defined distribution of S/CO ratios the median is a relevant measure, in addition to the mean. The difference between the median and the average of S/CO values may be an indicator of the skewness of the distribution curve. The value of this parameter  $\Delta(S/CO_{M-A})$  is expected to become higher with lower analytical sensitivity of the NAT system or lower virus concentration in the run control. It is recommended to use the Nelson rules<sup>21</sup> to identify deviations in the Levey Jennings trend analysis.

### ***Quantitative detection of HIV-1 RNA by viral load assays***

For monitoring the accuracy and precision in viral load assays one can use a Levey-Jennings QC chart for trend analysis.

#### ***Levey-Jennings QC chart.***

Test the run control at least 10 times during the reference period, apply log transformation on values expressed in IU/mL or copies/mL, estimate the geometric mean, standard deviation (SD) and its confidence interval (CI) as described below. [If Ct values are used no log transformation is required and confidence intervals can be calculated from the arithmetic mean and SD]. The Levey-Jennings chart is designed to identify individual aberrant values outside the 95% and 99% confidence intervals. With collecting additional data the chart characteristics may be updated.

The quantitative values for [HIV-1 RNA] are 'log normal' distributed.

- Calculate from each measurement the log(concentration) in IU/mL or copies/mL.
- Calculate mean and SD on these log values
- Take anti-log of the mean of log values, i.e. the geometric mean of the measurements in IU/mL or copies/mL.

Use table 4 to obtain Student-t-values belonging to the 95% and 99% CI for different number of observations (n). Calculate the log(95% and 99% CI) as follows:

- Log (99% Lower limit):  $\log(\text{Average}) - (99\% \text{ Student-t-Value} \times \log(\text{SD}))$
- Log (95% Lower limit):  $\log(\text{Average}) - (95\% \text{ Student-t-Value} \times \log(\text{SD}))$
- Log (95% Upper limit):  $\log(\text{Average}) + (95\% \text{ Student-t-Value} \times \log(\text{SD}))$
- Log (99% Upper limit):  $\log(\text{Average}) + (99\% \text{ Student-t-Value} \times \log(\text{SD}))$

**Table 4.** Relation of Student t value and numbers of runs (n) to calculate CI's.

Run (n)	t-value at 95% C.I.	t-value at 99% C.I.
10	2.306	3.355
20	2.101	2.878
30	2.048	2.763
infinite	1.960	2.576

Use the Nelson rules<sup>21</sup> to identify deviations in the Levey Jennings trend analysis.

### **Interpretation of test results on run control in Procleix Ultrio assay versions**

The expected frequency of S/CO values on P0064 ViraQ HIV-1 Check 125 control in the TMA assay as well as the interpretation of three categories of test result are shown in table 5.

The vast majority of S/CO values on the run control reach TMA signals in the (near) saturated range of the assay (between 8.0 and 15.0). Only a small fraction of TMA reactions on the run control are not yet complete and have S/CO values in the lower dynamic range of the assay (between 1.0 and 8.0). Repeatedly non-reactive results and a significant higher proportion of lower dynamic responses are indicative of reduced analytical sensitivity of the NAT system. A single event of a non-reactive result is however possible without deterioration of the test system and can be explained by Poisson distribution.



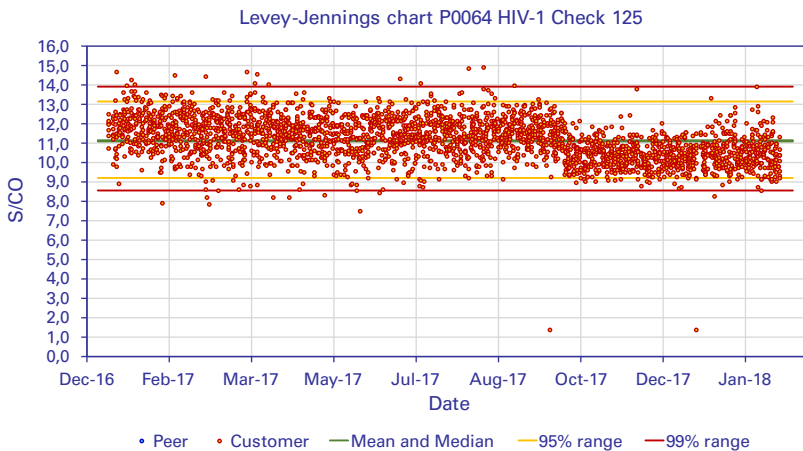
**Table 5.** Interpretation of a single TMA test result on P0064 ViraQ HIV-1 Check 125 in Procleix Ultrio assay versions and expected frequency of S/CO values in three ranges

Result	S/CO	Expected frequency per 1000	Interpretation
Reactive near saturated	>8.0	≥995	The test signal on the run control reaches values in the (near) saturated range of the TMA assay. This is an expected result.
Reactive lower dynamic	1.0–8.0	<5	The test signal on the run control is in the lower dynamic range of the assay because the TMA reaction is not yet complete. This is an expected result.
Non-reactive	<1.0	0 – 1	The test signal on the run control is below the cut-off. This is an unexpected result that should trigger an investigation of the technical performance of the test system.

**Monitoring performance of run control in Ultrio Elite assay**

Figure 2 and table 6 show Ultrio Elite performance data on P0064 ViraQ HIV-1 Check 125 obtained during 14 months of testing of the run control by one blood establishment using eight Panther instruments.

**Figure 2.** Levey-Jennings chart of P0064 ViraQ HIV-1 Check 125 control results in Grifols Ultrio Elite assay reported by one national blood organisation using eight Panther instruments. The average and median (green lines) and 95% and 99% CI (orange and red lines) are calculated as described in the text.



In 2904 Ultrio Elite test runs 5 S/CO values (0.17%) on the run control were below 8.0, of which 2 were just above the cut-off.

**Table 6.** Reproducibility of Ultrio Elite S/CO values on P0064 ViraQ HIV-1 Check 125 control observed during a 14 month observation period by one national blood service

n test runs	Median S/CO	Mean S/CO	$\Delta(S/CO_{M-A})$	S/CO Predictive interval	
				95%	99%
2904	11.13	11.13	0.00	9.2-13.1	8.6-13.9

The performance evaluation data can also be used to compare ViraQ run control batches, Ultrio Elite reagent lots or Panther instruments (table 7).

**Table 7.** Comparison of test results on different P0064 ViraQ HIV-1 Check 125 run control batches, Ultrio Elite reagent lots and Panther instruments during 14 months of testing by one national blood service

Item	Code	n	Median S/CO	St dev	Start date	End date
Run control product Batch	B4059-015	638	11,71	1,057	30-12-16	29-03-17
	B4059-016	635	11,44	0,992	30-03-17	28-06-17
	B4059-017	1631	10,77	1,037	29-06-17	11-02-18
Ultrio Elite Test Reagent Lot	154471	733	11,70	1,059	30-12-16	11-04-17
	159260	599	11,42	0,975	12-04-17	06-07-17
	171962	632	11,53	1,000	07-07-17	01-10-17
	180939	670	10,34	0,775	02-10-17	05-01-18
	186731	270	10,43	0,843	06-01-18	11-02-18
Panther instrument	1237	373	11,32	1,127	30-12-16	11-02-18
	1427	367	11,02	0,976	30-12-16	11-02-18
	1428	370	10,88	1,004	30-12-16	11-02-18
	1429	363	11,51	1,029	30-12-16	11-02-18
	1430	377	11,28	1,116	30-12-16	11-02-18
	1433	358	10,27	0,890	30-12-16	11-02-18
	1434	335	11,42	1,138	30-12-16	11-02-18
	1438	361	11,35	0,962	02-01-17	11-02-18

**Interpretation of test results on run control in quantitative NAT methods**

P0064 ViraQ HIV-1 Check 125 can be used as a quantitative run control in conjunction with the Hologic Aptima HIV-1 Quant tests and other viral load assays with a LOQ sufficiently below 125 copies/mL. Table 8 gives the expected frequency of three categories of results on the run control in viral load assays.

**Table 8.** Interpretation of a single quantitative NAT test result on P0064 ViraQ HIV-1 Check 125 control and expected frequency of viral load measurements above the lower limit of quantification (LOQ) of the current commercial real time PCR and TMA assays.

Result	HIV-cp/mL	Expected frequency	Interpretation
Reactive quantifiable	≥LOQ	>99%	This is an expected result.
Reactive unquantifiable	<LOQ	<1%	This is an unexpected result but is possible. An investigation of technical performance of the NAT system is recommended
Non-reactive undetectable	<CO	0%	This is an unexpected result. An investigation of technical performance of the NAT system is required

Repeatedly non-reactive or unquantifiable results are indicative of a significantly reduced analytical sensitivity of the NAT system. A single event of a test result below the LOQ is however possible without deterioration of the test system and can be explained by Poisson distribution

The linear range of the quantitative NAT methods tests starts at enough distance below the run control concentration of 125 copies/mL to expect quantifiable results (above the LOQ) in more than 99% of test runs<sup>5</sup>. The quantitative HIV-1 RNA assays report values in copies/mL. The HIV-1 RNA concentration (95%CI) of P0064 ViraQ HIV-1 Check control of 125 (95-163) copies/ (figure 1), 2.5- to 4.6-fold higher than the LOQ’s claimed by the manufacturers of the real time PCR and TMA assays (table 9).

**Table 9.** Distance of lower limit of quantification (LOQ) to concentration of P0064 ViraQ HIV-1 Check 125 control as reported in the package inserts of HIV-1 viral load assays of three manufacturers.

Manufacturer	NAT test	LOQ (cp/mL)	Factor (95%CI) <sup>#</sup>
Abbott	RealTime HIV-1	40	3.1 (2.5 – 4.1)
Roche Molecular systems	HIV-1 Cobas 6800/8800	48	2.6 (2.0 – 3.4)
Hologic	Aptima HIV-1 Quant	30	4.6 (4.2 – 5.5)

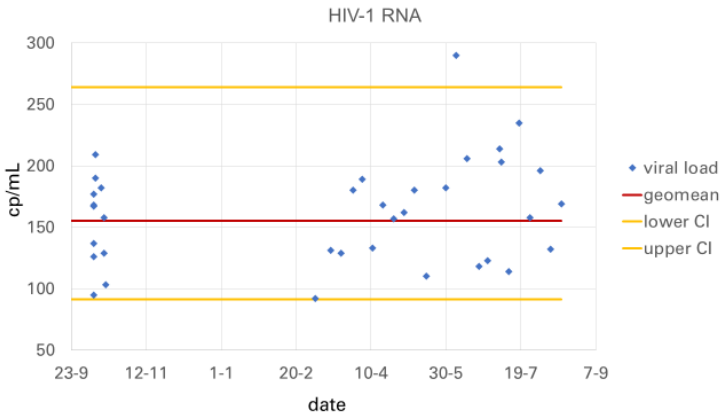
<sup>#</sup>Factor between Concentration of HIV-1 RNA in copies/mL of P0064 ViraQ HIV-1 Check 125 and LOQ; 95% CI’s derived from uncertainty in calibration of P0064 ViraQ P0063 Check 125 control.

One should be careful with comparing the copies/mL levels in table 9 because different methods and standards have been used for calibration of the run control and (calibrators of) the NAT systems.

### Monitoring performance of quantitative NAT methods on run control

For the identification of aberrant quantitative results log (viral load) values should be recorded in a Levey-Jennings chart to visualise trends over time. The Nelson rules<sup>21</sup> provide guidance on the interpretation of results outside the 95% or 99% confidence intervals. An example is given in figure 3 showing data points of Aptima Quant test runs on P0064 ViraQ HIV-1 Check 125 control in a Levey-Jennings scatter plot. Testing of P0064 ViraQ HIV-1 Check 125 control in 34 test runs of the Aptima HIV Quant assay gave a geometric mean value (95% CI) of 155 (91-264) copies/mL comparable to the estimated concentration of 125 (95-163) IU/mL in the run control<sup>17</sup>.

**Figure 3.** Reproducibility of Hologic Aptima HIV Quant test runs on P0064 ViraQ HIV-1 Check 125 control presented in a Levey-Jennings chart.



The distance from the geometric mean viral load (green line in graph) represents the deviation from the expected TMA response level on the run control. The orange lines represent the 95% CI.

One can use the quantitative results on the run control for comparison of different experimental conditions, such as different laboratories, NAT reagent batches or instruments. Since the concentration of P0064 HIV-1 Check 125 is just above the Poisson detection endpoint range of the quantitative NAT methods, lower reported copies/mL values on the run control or reduced analytical sensitivity of the test system may coincide with an increased standard deviation<sup>17</sup>.

## Limitations

- P0064 ViraQ HIV-1 Check 125 Control cannot be used to evaluate the analytical or diagnostic sensitivity of NAT blood screening assays (although a significant reduction of analytical sensitivity of the NAT system can become apparent with repeated occurrence of non-reactive or unquantifiable results).
- P0064 ViraQ HIV-1 Check 125 Control must not be substituted for the mandatory controls or calibrators provided with NAT test kits for calculating the cut-off and/or criteria for releasing test results.
- The Poisson distribution in samples with low HIV-1 concentrations cannot guarantee that 100% reactive results will be found on P0064 ViraQ HIV-1 Check 125 Control in NAT blood screening assays. Therefore the response values on the run controls should not be used for a decision to accept or reject the test run.
- The expected distributions of assay response values on P0064 ViraQ HIV-1 Check 125 Control that are presented in this package insert were based on evaluation studies involving a limited number of tests and NAT reagent lots. Therefore it cannot be guaranteed that different results will be found on other assay versions or NAT reagent lots.
- The parameter  $\Delta(S/CO_{M-A})$  is an indicator of the skewness of the distribution curve of S/CO values and may be a performance indicator for the analytical sensitivity of Ultrio assay versions and reagent lots. However a threshold value above which a deterioration of the test system is predicted cannot be given.
- P0064 ViraQ HIV-1 Check 125 should not be used for establishing accuracy of quantitative NAT results expressed in IU/mL. For this purpose only a dilution of the current WHO International Standard can be used.
- More quantitative data need to be collected in the viral load assays to confirm the suitability of P0064 HIV-1 Check 125 control for these methods and to ensure that the proportion of unquantifiable results is less than 1%.

## References

1. Grabarczyk P, van Drimmelen H, Kopacz A, Gdowska J, Liszewski G, Piotrowski D, Górka J, Kuśmierczyk J, Candotti D, Lętowska M, Lelie N, Brojer E. Head-to-head comparison of two transcription-mediated amplification assay versions for detection of hepatitis B virus, hepatitis C virus, and human immunodeficiency virus Type 1 in blood donors. *Transfusion*. 2013; 53:2512-2524.
2. Assal A, Barlet V, Deschaseaux M, Dupont I, Gallian P, Guitton C, Morel P, David B, and De Micco P. Comparison of the analytical and operational performance of two viral nucleic acid test blood screening systems: Procleix Tigris and cobas s 201. *Transfusion* 2009; 49:289-300.
3. Koppelman M, Assal A, Chudy M, Torres P, de Villaescusa RG, Reesink HW, Lelie PN, Cuypers HT. Multi-center performance evaluation of a transcription-mediated amplification assay for screening of human immunodeficiency virus-1 RNA, hepatitis C virus RNA, and hepatitis B virus DNA in blood donations. *Transfusion* 2005; 45:1258-66.
4. Grabarczyk P, Koppelman M, Boland F, Saulea S, Fabra C, Cambie G, O'Riordan K, Van Drimmelen H, Vermeulen M, O'Riordan J, Lelie N. Inclusion of human immunodeficiency virus Type 2 (HIV-2) in a multiplex transcription-mediated amplification assay does not affect detection of HIV-1 and hepatitis B and C virus genotypes: a multicenter performance evaluation study. *Transfusion* 2015; 55:2246-55.
5. Lelie PN, Van Drimmelen AAJ. Positioning of ViraQ Check and Trend Controls compatible with analytical sensitivity of NAT assays. VR4059. [www.bioqcontrol.com](http://www.bioqcontrol.com)
6. Lelie PN, Van Drimmelen AAJ. Preparation of inactivated secondary viral standards: Safety assessment of quality control samples for viral serology and NAT assays in blood screening laboratories.CE4006. [www.bioqcontrol.com](http://www.bioqcontrol.com)
7. Lelie PN, Reesink HW, Lucas CJ. Inactivation of 12 viruses by heating steps applied during manufacture of a hepatitis B vaccine. *J Med Virol*. 1987;23:297-301.
8. Tersmette M, de Goede RE, Over J, de Jonge E, Radema H, Lucas CJ, Huisman HG, Miedema F. Thermal inactivation of human immunodeficiency virus in lyophilised blood products evaluated by ID50 titrations. *Vox Sang*. 1986;51:239-43
9. Bruzzone B, Bisio F, Caligiuri P, Mboundou FA, Nigro N, Sticchi L, Ventura A, Saladini F, Zazzi M, Icardi G, Viscoli C. Discordances with HIV-1 RNA quantitative determinations by three commercial assays in Pointe Noire, Republic of Congo. *J Virol Methods*. 2014;203:102-6
10. Sauné K, Delaugerre C, Raymond S, Nicot F, Boineau J, Pasquier C, Izopet J. Analytical sensitivity of three real-time PCR assays for measuring subtype B HIV-1 RNA. *J Clin Virol*. 2013;57:80-3.
11. Pyne MT, Wilson A, Hillyard DR. Large-scale comparison of Roche Cobas AmpliPrep/Cobas TaqMan and Abbott RealTime HIV assays. *J Virol Methods*. 2012 ;184:106-8.
12. Collins ML, Zayati C, Detmer JJ, Daly B, Kolberg JA, Cha TA, Irvine BD, Tucker J, Urdea MS. Preparation and characterization of RNA standards for use in quantitative branched DNA hybridization assays. *Anal Biochem*. 1995 20; 226:120-9.
13. Lelie PN, Van Drimmelen AAJ, Cuypers HTM, Best SJ, Stramer Hyland SL C, J.- Allain P, Moncharmont P, Defer C, Nubling CM, Glauser A, da Silva Cardoso M, -F. Viret J, Lankinen M, Grillner L, Wirthmuller U, Coste J, Schottstedt V, Masecar B. and E.M. Dax. Sensitivity of HCV-RNA and HIV-RNA blood screening assays. *Transfusion*. 2002,42:527-36.

14. Lelie PN, Van Drimmelen AAJ. Calibration of native and inactivated viral standards and traceability to viral nucleic acid copies and International Units. VR4060, [www.bioqcontrol.com](http://www.bioqcontrol.com)
15. Holmes H, Davis C, Heath A, Hewlett I and Lelie PN. An international collaborative study to establish the 1st International Standard for HIV-1-RNA for use in Nucleic Acid-Based Techniques. *J. Virol. Methods* 2001, 92: 141-150
16. C. Davis, A. Heath, S. Best, I. Hewlett, N. Lelie, R. Schuurman, H. Holmes Calibration of HIV-1 working reagents for nucleic acid amplification techniques against the 1st international standard for HIV-1 RNA. *J of Virol Methods* 2003;107:37-44.
17. Van Drimmelen AAJ, Lelie PN. Performance evaluation of ViraQ run controls for HBV, HCV and HIV-1 detection in different NAT assays. VR4061. [www.bioqcontrol.com](http://www.bioqcontrol.com)
18. Van Drimmelen AAJ, Lelie PN. Stability of ViraQ run controls for NAT. VR4058. [www.bioqcontrol.com](http://www.bioqcontrol.com)
19. Centers for Disease Control (CDC). Update: Universal precautions for prevention of transmission of human immunodeficiency virus, hepatitis B virus, and other blood borne pathogens in health-care settings. *MMWR* 1988; 37:377-388.
20. Centers for Disease Control (CDC). Guidelines for prevention of transmission of human immunodeficiency virus and hepatitis B virus to health-care and public-safety workers. *MMWR* 1989; 38(S-6): 1-36.
21. Nelson LS, "The Shewhart Control Chart—Tests for Special Causes". *Journal of Quality Technology* 1984;16, no. 4: 238-239  
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