

P0069 ViraQ HBV Trend 25





The kit insert contains a detailed protocol and should be read carefully before testing the run control to ensure optimal performance



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Intended Use

P0069 ViraQ HBV Trend 25 is intended to be used as external trend control for hepatitis B virus (HBV)-DNA detection by the multiplex transcription mediated amplification (TMA) assays on the automated nucleic acid amplification technology (NAT) platforms defined in Table 1. The trend control helps laboratories to ensure that HBV is detected with sufficient analytical sensitivity by consecutive reagent lots of the Procleix Ultrio assay versions and by each of the Tigris or Panther instruments in use. The trend control can be used in daily test runs to continuously monitor NAT performance over time or tested occasionally in multiple replicates in one test run for:

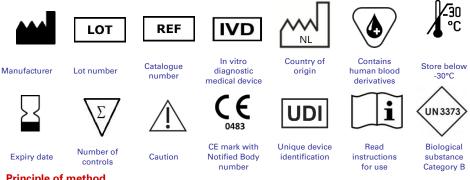
- acceptance (transport integrity) testing of TMA reagent lots
- installation qualification of instruments
- training of technicians.

Table 1. Assays and platforms covered by P0069 ViraQ HBV Trend control

Assays (manufacturer)	Platform	Test environment	
Procleix Ultrio Plus® (Grifols)	Procleix Tigris [®]	Diago di serre animar	
Procleix Ultrio Elite® (Grifols)	Procleix Panther®	 Blood screening 	

P0069 ViraQ HBV Trend 25 should not be used to replace the internal controls or calibrators in the test kits. The test result on the run control should not be used to reject the run or delay the release of test results on donor samples.

Key to Symbols Used



Principle of method

P0069 ViraQ HBV Trend 25 control has been formulated to mimic natural plasma specimens with a low HBV-DNA concentration. After thawing the trend control tubes are ready for use and can be placed at random positions in sample racks on the NAT platforms. The trend control contains 25 copies/mL of HBV-DNA (equivalent to 4.7 International Units (IU)/mL) and has been designed to ensure sufficient analytical sensitivity of transcription mediated amplification (TMA) tests in blood screening laboratories. The HBV-DNA concentration in the run control has been set near the 95% lower limit of detection (LOD) of the Ultrio Plus and Elite assays (table 2)¹⁻⁵. P0069 ViraQ HBV Trend 25 Control enables laboratories to be alerted in case of a reduction of analytical sensitivity of NAT instruments or reagent lots and to identify changes in TMA performance over time. The run control is a dilution of the S0043 HBV-RNA genotype A2 standard, prepared by heat-inactivation of a pool of HBsAg plasma units from the same

donor⁶⁻⁸. The plasma matrix in which the run control is diluted is manufactured from plasma units that tested negative for all relevant markers of blood borne viruses. The S0043 HBV standard has been calibrated in copies/mL and IU/mL against the Viral Quality Control (VQC)-Sanguin, Eurohep and World Health Organization (WHO) International Standards (figure 1). The low concentration of HBV genotype A in the run control is representative for HBV Genotypes A to H that are prevalent in different geographical regions of the world (and that are detected with similar analytical sensitivity by the above mentioned TMA assays)^{4,9}. A positive result on the trend control indicates that the NAT method has been performed with sufficient analytical sensitivity. A higher than expected proportion of non-reactive or weakly reactive results is indicative of reduced analytical sensitivity of the NAT system and should trigger investigation of the technical performance of the assay. The run control generates sample to cut-off (S/CO) ratios in the Procleix Ultrio assay versions, Statistical analysis of these assay response values generated over a certain period of time allows for comparison of analytical performance of NAT reagent lots and laboratory instruments. The trend control can also be used in multiple replicates in the same test run to ensure that TMA reagents or instruments fulfil the minimum requirements for analytical sensitivity before they are accepted for routine blood screening.

standard	panel	NAT method	n	50% LOD (CI) cp/mL	95% LOD (CI) cp/mL
	P0031	Ultrio Plus	24	6.6 (2.7-17.4)	64 .2 (22.4-1099)
S0043 BioQ HBV-DNA genotype A inact.	P0031	Ultrio Elite	25	5.7 (4.0-8.2)	40.8 (24.3-91.7)
3, p	P0031	Ultrio Plus/Elite	49	7.6 (5.9-9.5)^	33.3 (23.8-56.4)^
S0011 VQC-Sanquin HBV-DNA genotype A	P0007	Ultrio Plus	48	4.8 (3.7-6.2)	38.8 (25.6-68.5)
	P0007	Ultrio Elite	74	3.4 (2.3-4.8)	43.2 (24.8-98.0)
	P0007	Ultrio Plus/Elite	122	4.3 (2,9-6,1)^	35.4 (20,6-87,8)^
S0010 Eurohep HBV-	P0001	Ultrio Plus	96	3.6 (2.9-4.4)	40.4 (29.2-60.2)
DNA genotype A	P0001	Ultrio Elite	24	7.9 (5.5-11.2)	49.1 (29.4-116)
WHO HBV-DNA	P0023	Ultrio Plus	303	4.4 (3.3-5.9)	28.4 (18.0-57.7)
97/750#	P0023	Ultrio Elite	252	4.4 (3.6-5.4)	30.9 (22.4-47.4)

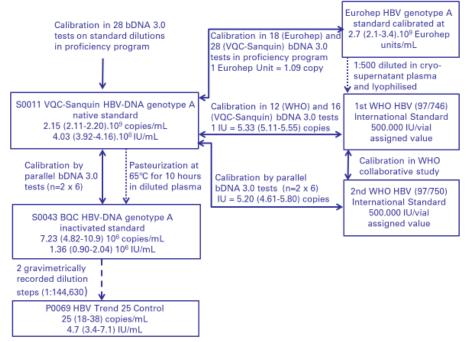
 Table 2. Detection limits on native and inactivated HBV standard dilution panels in Procleix
 Ultrio assay versions

^ probit analysis without two lowest concentrations in dilution panel #1 IU = 5.33 copies

Traceability to HBV-DNA copies and International Units

Figure 1 shows the traceability chain between the ViraQ run control, the Bio Quality Control (BQC) standard, VQC-Sanquin standard, the Eurohep standard and the 1st and 2nd WHO 97/746 and 96/750 International Standards for HBV-DNA. The inactivated S0043 HBV-DNA standard (used for preparation of the P0069 ViraQ trend control) has been calibrated in copies/mL by replicate testing in the Siemens Versant bDNA 3.0 assay¹⁰ against the historically established S0011 VQC-Sanquin HBV-DNA genotype A standard¹¹. The VQC-Sanquin HBV-DNA genotype A standard has been calibrated at 5.33 (5.11-5.55) and 5.20 (4.61-5.80) copies per IU against the first and second WHO HBV-DNA (97/746 and 97/750) standards respectively in two experiments¹². It must be emphasized that this conversion factor from copies to IU values has not been confirmed for the later 3rd WHO 10/264 replacement standard. The copy number assigned to the VQC-Sanquin standard was found to be comparable to that of the Eurohep standard¹³ used for preparation of the WHO standards¹⁴. The accurate calibration of the VQC-Sanquin and the inactivated BQC standard against the WHO and Eurohep standards in IU/mL and in copies/mL has been confirmed in analytical sensitivity studies of the Grifols Procleix TMA and Roche cobas MPX assays^{4,12}. The BQC manufacturing and quality control procedures guarantee consistent virus concentrations in consecutive ViraQ HBV Trend 25 batches¹⁵. The inactivated BQC HBV genotype A standard is available in sufficient supply to ensure batch to batch consistency of ViraQ trend controls for a prolonged period of time.





Stability of HBV standards and run control

The long term stability of the liquid frozen S0043 HBV standard stored at \leq 65°C has been firmly established¹⁶; hence the stock solution from which the trend control is prepared has shown to be stable in the BioQControl storage facilities. Real time stability experiments using quantitative NAT assays showed no degradation of HBV-DNA in P0065 ViraQ HBV Check 125 control when stored at -30°C¹⁶. Hence, it can be guaranteed that also the P0069 trend control is still functional and should generate a reactivity rate near 95% when stored at -30°C and used before the expiration date (two years after preparation of the run control batch)^{15,16}.

Kit contents (materials provided)

The run control contains human plasma without preservatives and is provided in three formats as detailed in Table 3.

P0069/01 and P0069/02 are intended to accommodate blood screening laboratories. To facilitate automation the run control is presented in a polypropylene tube with screw cap comparable in size to vacutainer tubes used for donor sample collection. The tube label has a barcode identifying the product, sequential batch number and marker HBV. The barcode can be read by the automated NAT systems.

Cat. Code	UDI code	Quantity run control	Size vials	packing	
P0069/01 P0069/02	8718719830695 8718719830289	60 x 1.5 mL 10 x 1.5 mL	10 mL 10 mL	60 vials in rack/box Plastic zip bag	
10000/02	0,10,10000200	10 X HO HE	10 IIIE	i laotio Lip bag	

Table 3. Description of kit formats and contents

Materials required but not supplied

The test kits and liquid handling devices provided by the NAT manufacturer as specified in Table 1.

Storage instructions

The trend controls should be stored at or below -30°C for a maximum of two years¹⁶. Once thawed the run control samples should be used within 8 hours. During this period, when not in use, store sample at 2-8°C¹⁷. Do not refreeze the controls after thawing to prevent formation of cryoprecipitates. Any control sample that appears cloudy or contains precipitates after thawing and mixing should be discarded.

Warning and precautions

Although P0069 ViraQ HBV Trend 25 contains inactivated HBV particles⁶⁻⁸ the plasma may still be potentially bio-hazardous. The matrix is prepared from human blood plasma that tested negative for blood borne viruses (HBV-DNA, HCV-RNA, HIV-RNA, HBsAg, anti-HBc, anti-HBs, anti-HIV, anti-HCV and anti-Treponema *pallidum*). No test method can offer complete assurance that products derived from human blood cannot transmit (unknown) infectious agents. The run control should only be used by trained laboratory workers who are aware of the potential risk of infectious agents in human plasma samples and take the necessary precautions. Observe the universal precautions for prevention of transmission of infectious agents when handling these materials^{17,18}.

- Do not pipette by mouth.
- Use personal protective equipment, including lab coats, gloves and safety glasses.
- Do not eat, drink or smoke in areas where the run controls is handled.
- Disinfect spills using a 0.5% hypochlorite solution (1:10 v/v household bleach) or equivalent disinfectant.
- Dispose unused or spilled materials according to the normal practices for biological waste disposal in your institution.
- If precipitates are visible, mix the run controls for 2 minutes thoroughly.
- Once thawed, do not re-freeze and thaw the run control samples to avoid formation of cryoprecipitates that could alter reactivity or cause pipetting errors in the automated sampling systems.
- Store run controls in an upright position.

Reagent preparation

- Thaw the run control quickly in a water bath at 37°C.
- Mix gently during thawing until contents are just thawed.
- Immediately after thawing remove the run control tube from the water bath.
- Vortex the run control.
- Give a short spin in a centrifuge to remove liquid before releasing screw cap from vial.
- Minimise the time period from thawing until usage of the control samples.
- Use within 8 hours after thawing
- After thawing when not in use: store at 2-8°C

Test procedure and calculations

The trend control should be tested in a manner identical to that of clinical specimens and the result be calculated according to the instructions for use of the NAT procedure.

The following sections in this package insert provide guidance on interpretation and analysis of test results on P0069 ViraQ HBV Trend 25. The statistical evaluation methods were developed by BioQ Control and not reviewed nor approved by the manufacturer of the Ultrio assay versions

Qualitative detection of HBV-DNA in Procleix Ultrio versions

The results of the Procleix Ultrio Plus and Ultrio Elite assays are expressed as a sample to cut-off ratio (S/CO). P0069 ViraQ HBV Trend 25 Control should react positive in approximately 90% to 95% of TMA test runs. Approximately 90% of test results on the trend control are expected in the (near) saturated range of the TMA assay with S/CO values equal to or above 12.0. Approximately 2% to 10% of test results are expected in the dynamic range of the TMA assay with S/CO rations below 12.0 (see interpretation of test results below)¹⁵. A Levey-Jennings QC chart can be used to monitor the performance of the Ultrio assay versions on the run control.

Levey-Jennings QC chart.

The S/CO responses on ViraQ HBV Trend 50 in the Ultrio Plus and Elite assay versions are not normally distributed, also not after transformation of the S/CO ratios. For developing a Levey-Jennings QC chart a distribution-free approach can be taken whereby the ranges containing 95% and 99% of the data are calculated. For this purpose the 0.5% and 99.5% percentiles are calculated for the 99% predictive interval, and the 2.5% and 97.5% percentiles for the 95% predictive interval. In case the total dataset does not contain sufficient data, values just outside the observed range can be presented in order to allow for a graphical presentation. For this non-defined distribution of S/CO ratios the median is a relevant measure, in addition to the mean. The difference between the median and the average of S/CO values is an indicator of the skewness of the distribution curve. The value of this parameter Δ (S/CO_{M-A}) is expected to become higher with lower analytical sensitivity of the NAT system or lower virus concentration in the run control. It is recommended to use the Nelson rules¹⁹ to identify deviations in the Levey Jennings trend analysis.

Interpretation of test results on trend control in Procleix Ultrio assay versions

The expected frequency of S/CO values on P0069 ViraQ HBV Trend 25 control in the below cut-off, the lower dynamic and the (near) saturated range of the TMA assay as well as the interpretation of these categories of test result are shown in table 4. The majority of S/CO values on the run control reach near saturated TMA response levels and are found between 12.0 and 15.0 (figure 2). Approximately 5 (2-9)% of TMA reactions on the trend control are not yet complete and have S/CO values in the lower dynamic range of the assay (between 1.0 and 12.0). In a one month observation period of 190 Ultrio Elite test runs the overall proportion of reactive results on P0069 HBV Trend 25 control was 92.6% as compared to 98.3% on P0154 HBV Trend 50 control tested in parallel (table 5)¹⁶. The latter trend control containing 50 (instead of 25) copies/mL was tested during 4 years with reactivity rates varying between 97.2% and 100% between TMA reagent lots and trend control batch combinations)¹⁵.

 Table 4.
 Interpretation of a single TMA test result on P0069 ViraQ HBV Trend 25 in Procleix Ultrio assay versions and expected frequency of S/CO values in three ranges

ranges			
Result	S/CO	Expected frequency per 1000 [#]	Interpretation
Reactive (near) saturated	>12.0	736 – 1000	The test signal on the trend control reaches values in the (near) saturated range of the TMA assay. This is an expected result.
Reactive lower dynamic	1.0–12.0	24 –92	The test signal on the run control is in the lower dynamic range of the assay because the TMA reaction is not yet complete. This is an expected result.
Non- reactive	<1.0	35-112	The test signal on the run control is below the cut-off. This is an expected result
#			

*95% confidence limits found in 190 Ultrio Elite test runs

Table 5. Comparison of reactivity rate on HBV Trend Control (TC) batches of 50 and 25 copies/mL in two Ultrio Elite (UE) reagent batches.

Ultrio	P0154 ViraQ HBV 50			P006	′ 25	
Elite batch	Trend control batch	reactive/n	%	Trend control batch	reactive/n	%
UE1	TC4	427/434	98.5%	TC1	54/58	93.1%
UE2	TC4	138/141	97.9%	TC1	122/132	92.4%
UE all	TC4	565/575	98.3%	TC1	176/190	92.6%

Monitoring performance of Procleix Ultrio assay versions on trend control

Figure 2 shows Ultrio Elite performance data on P0069 ViraQ HBV Trend 25 obtained during one month of testing by one national blood service in a Levey-Jennings QC Chart. Table 6 presents the same Ultrio Elite data as presented in Figure 2.

Figure 2. Levey-Jennings chart of P0069 ViraQ HBV Trend 25 Control results in Grifols Ultrio Elite assay reported by one national blood organisation using four Panther instruments. The average and median (green lines) and 95% and 99% predictive intervals (orange and red lines) are calculated as described in the text.

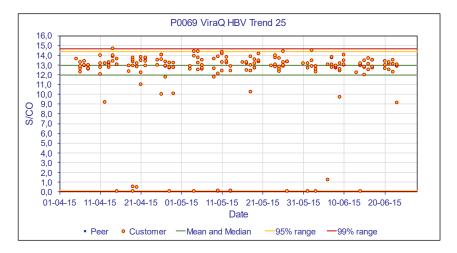


Table 6. Reproducibility of Ultrio Elite S/CO values on P0069 ViraQ HBV Trend 25 control

n test	Median	Average	∆(S/CO _{M-A})	S/CO Predi	ctive interval
runs	S/CO	S/CO	∆(5/COM-A)	95%	99%
190	13.00	12.02	0.98	0.09 – 14.0	0.07 – 14.4

The difference between the median and the average of S/CO values can be used as an indicator of the skewness of the distribution curve and the analytical sensitivity of the NAT system (table 6). Based on the available results one may conclude that if Δ (S/CO_{M-A}) is below 1.6 the system is properly functioning¹⁵. Data on another ViraQ HBV trend control of 50 copies/mL showed that the presence of a higher proportion of non-reactive results coincides with high values of Δ (S/CO_{M-A}), thereby confirming its ability to be a trend indicator for analytical performance of the TMA assay. An alert threshold value for this parameter that is indicative for poor NAT performance cannot be given with the available data.

The parameter Δ (S/CO_{M-A}) can also be applied to compare other experimental conditions such as the TMA reagent lot, the ViraQ trend control batch or the testing robot (Tigris or Panther). Hence, if the reagent lot performance indicator Δ (S/CO_{M-A}) has an outlier value it could be used as an alert signal for checking technical performance of that particular TMA reagent lot or trend control batch.

Acceptance testing of NAT system component using trend control

P0069 ViraQ HBV Trend 25 can also be used for acceptance testing of a new TMA reagent lot, a new trend control batch, a reagent transport integrity check, a (re)-installation qualification of a Panther or Tigris instrument or training of an operator. For these applications it is recommended to test 20 vials of the trend control in one Ultrio Plus or Elite test run. The reagent batch, instrument or operator performance is approved when at least 17/20 (85%) of tests are reactive and the median S/CO value is above 12.7. If either one of these criteria is not fulfilled it is recommended to repeat the acceptance test procedure in another test run. If in the repeat test either one of these criteria is again not fulfilled further investigation of the performance of the reagent lot or instrument is recommended. These acceptance criteria were established by a simulation study with sliding sets of 20 sequential results out of a data base of only 190 Elite test runs¹⁵. A preliminary decision algorithm for accepting the NAT system component is summarized in table 7.

Table 7. Decision algorithm and criteria for acceptance of reagents, instruments or operators by replicate testing of 20 vials of P0069 ViraQ HBV Trend 25 control in one Procleix Ultrio (Plus or Elite) assay run

Acceptar	nce criteria	_ Expected	
reactivity rate ≥17/20 (85%)	Median S/CO ≥12.7	frequency	Decision
ОК ОК		>95%	Accept
either one of criteria r	not fulfilled on initial test	<5%	Repeat acceptance test protocol
either one of criteria n	ot fulfilled on repeat test	<0.25%	Initiate root cause analysis

Limitations

- P0069 ViraQ HBV Trend 25 Control cannot be used to determine the analytical or diagnostic sensitivity of NAT blood screening assays (although changes in analytical sensitivity of the NAT system can become apparent with the trend control).
- P0069 ViraQ HBV Trend 25 Control must not be substituted for the mandatory controls or calibrators provided with NAT test kits for calculating the cut-off and/or criteria for releasing test results.
- A single nonreactive test result on P0069 ViraQ HBV Trend 25 Control cannot be used to invalidate a test run. The Poisson distribution in samples with low HBV concentrations cannot guarantee that the response values are reproducible. Therefore the trend control cannot be used for a decision to accept or reject a test run.
- The expected distribution of assay response values on P0069 ViraQ HBV Trend 25 Control that is presented in this package insert was based on an evaluation study involving a limited number of tests and NAT reagent batches. Therefore it cannot be guaranteed that different results will be found on other assay versions or NAT reagent lots.
- The parameter ∆(S/CO_{M-A}) as performance indicator of Ultrio Plus and Elite assays and the proposed threshold value of 1.60 above which a deterioration of the test system is possible needs to be further evaluated and confirmed in post-market performance follow up studies.
- The decision algorithm for acceptance testing of NAT system components was based on testing of a limited number of Ultrio Elite reagent lots. The validity of the acceptance criteria need to be confirmed by testing more Ultrio Elite reagent lots on the P0069 HBV trend control in multiple replicates.

References

- Grabarczyk P, van Drimmelen H, Kopacz A, Gdowska J, Liszewski G, Piotrowski D, Górska J, Kuśmierczyk J, Candotti D, Lętowska M, Lelie N, Brojer E. Head-tohead comparison of two transcription-mediated amplification assay versions for detection of hepatitis B virus, hepatitis C virus, and human immunodeficiency virus Type 1 in blood donors. Transfusion. 2013; 53:2512-2524.
- Assal A, Barlet V, Deschaseaux M, Dupont I, Gallian P, Guitton C, Morel P, David B, and De Micco P. Comparison of the analytical and operational performance of two viral nucleic acid test blood screening systems: Procleix Tigris and cobas s 201. Transfusion 2009; 49:289-300.
- Koppelman M, Assal A, Chudy M, Torres P, de Villaescusa RG, Reesink HW, Lelie PN, Cuypers HT. Multi-center performance evaluation of a transcriptionmediated amplification assay for screening of human immunodeficiency virus-1 RNA, hepatitis C virus RNA, and hepatitis B virus DNA in blood donations. Transfusion 2005; 45:1258-66.
- Grabarczyk P, Koppelman M, Boland F, Sauleda S, Fabra C, Cambie G, O'Riordan K, Van Drimmelen H, Vermeulen M, O'Riordan J, Lelie N. Inclusion of human immunodeficiency virus Type 2 (HIV-2) in a multiplex transcription-mediated amplification assay does not affect detection of HIV-1 and hepatitis B and C virus genotypes: a multicenter performance evaluation study. Transfusion 2015; 55:2246-55.
- Lelie PN, Van Drimmelen AAJ. Positioning of ViraQ Check and Trend Controls compatible with analytical sensitivity of NAT assays. VR4059. www.bioqcontrol.com
- Lelie PN. Van Drimmelen AAJ. Preparation of inactivated secondary viral standards: Safety assessment of quality control samples for viral serology and NAT assays in blood screening laboratories.CE4006. www.bioqcontrol.com
- Lelie PN, Reesink HW, Niessen J, Brotman B, Prince AM. Inactivation of 10¹⁵ chimpanzee-infectious doses of hepatitis B virus during preparation of a heatinactivated hepatitis B vaccine. J Med Virol. 1987 Nov;23(3):289-95
- Lelie PN, Reesink HW, Lucas CJ. Inactivation of 12 viruses by heating steps applied during manufacture of a hepatitis B vaccine. J Med Virol. 1987;23:297-301.
- 9. Chudy M, Hanschmann KM, Kress J, Nick S, Campos R, Wend U, Gerlich W, Nübling CM.First WHO International Reference Panel containing hepatitis B virus genotypes A-G for assays of the viral DNA. J Clin Virol. 2012;55:303-9
- Collins ML, Zayati C, Detmer JJ, Daly B, Kolberg JA, Cha TA, Irvine BD, Tucker J, Urdea MS. Preparation and characterization of RNA standards for use in quantitative branched DNA hybridization assays. Anal Biochem. 1995 20; 226:120-9.
- Lelie PN, Van Drimmelen AAJ, Cuypers HTM, Best SJ, Stramer Hyland SL C, J.-Allain P, Moncharmont P, Defer C, Nubling CM, Glauser A, da Silva Cardoso M, -F. Viret J, Lankinen M, Grillner L, Wirthmuller U, Coste J, Schottstedt V, Masecar B. and E.M. Dax. Sensitivity of HCV-RNA and HIV-RNA blood screening assays. Transfusion. 2002,42:527-36.
- Lelie PN, Van Drimmelen AAJ. Calibration of native and inactivated viral standards and traceability to viral nucleic acid copies and International Units. VR4060, www.bioqcontrol.com
- Heermann KH, Gerlich WH, Chudy M, Schaefer S, Thomssen R.J. Quantitative detection of hepatitis B virus DNA in two international reference plasma preparations. Eurohep Pathobiology Group. Clin Microbiol. 1999;37:68-73.

- Saldanha J, Gerlich W, Lelie N, Dawson P, Heermann K, Heath A; WHO Collaborative Study Group. An international collaborative study to establish a World Health Organization international standard for hepatitis B virus DNA nucleic acid amplification techniques. Vox Sang. 2001;80:63-71.
- Van Drimmelen AAJ, Lelie PN. Performance evaluation of ViraQ run controls for HBV, HCV and HIV-1 detection in different NAT assays. VR4061. www.bioqcontrol.com
- 16. Van Drimmelen AAJ, Lelie PN. Stability of ViraQ run controls for NAT. VR4058. www.bioqcontrol.com
- 17. Centers for Disease Control (CDC). Update: Universal precautions for prevention of transmission of human immunodeficiency virus, hepatitis B virus, and other blood borne pathogens in health-care settings. MMWR 1988; 37:377-388.
- Centers for Disease Control (CDC). Guidelines for prevention of transmission of human immunodeficiency virus and hepatitis B virus to health-care and publicsafety workers. MMWR 1989; 38(S-6): 1-36.
- Nelson LS, "The Shewhart Control Chart—Tests for Special Causes". Journal of Quality Technology 1984;16, no. 4: 238-239 https://doi.org/10.1080/00224065.1984.11978921



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